

Visium CytAssist Spatial Gene and Protein Expression – FFPE Tissue Preparation Guide

Introduction

The Visium CytAssist Spatial Gene and Protein Expression assay is designed to analyze mRNA and protein expression in tissue sections derived from formalin fixed & paraffin embedded (FFPE) tissue samples. A single CytAssist run accommodates up to two tissue slides (tissue placed on a standard glass slide) as sample input. Proper tissue handling and preparation techniques preserve the morphological quality of the tissue sections and the integrity of mRNA transcripts. Maintaining tissue adhesion, high-quality RNA, and protein epitope antigenicity are critical to assay performance.

The Tissue Preparation Guide provides guidance on:

- Best practices for handling tissue samples and tissue slides before and after sectioning.
- Removing hardset coverslips from archived Hematoxylin & Eosin (H&E) stained tissue slides.
- Sectioning of tissue blocks and placement of sections on blank slides.
- Performing RNA quality assessment of FFPE tissue blocks or archived tissue sections on tissue slides.

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Additional Guidance

This protocol is compatible with most tissue types. Modifications to this protocol (e.g. section flotation time and water bath temperature) may be required for the preparation of certain tissue blocks and difficult tissue types, such as breast, colon, skin, and lungs. Refer to the 10x Genomics Support website for additional resources, including a list of tissues tested.

The tissue slides prepared using this Tissue Preparation Guide can be used with:

- Visium CytAssist Spatial Gene and Protein Expression – Deparaffinization, H&E Staining, Imaging & Decrosslinking for FFPE Tissue (CG000658)
- Visium CytAssist Spatial Gene and Protein Expression – Deparaffinization, Decrosslinking, IF Staining & Imaging for FFPE Tissue (CG000659)

Document Navigation

- If archived or freshly placed tissue sections are larger than the Visium Capture Area (Refer to [Determining Allowable Area](#) for dimensions), define an area of interest. Refer to [Optional Area of Interest Selection for Large Tissue Sections](#) on page 8 for more information.
- If working with hardset coverslipped, H&E stained archived slides, protocol steps begin on page 13.
- If working with FFPE tissue blocks, protocol steps begin on page 20.

10x Genomics Accessories

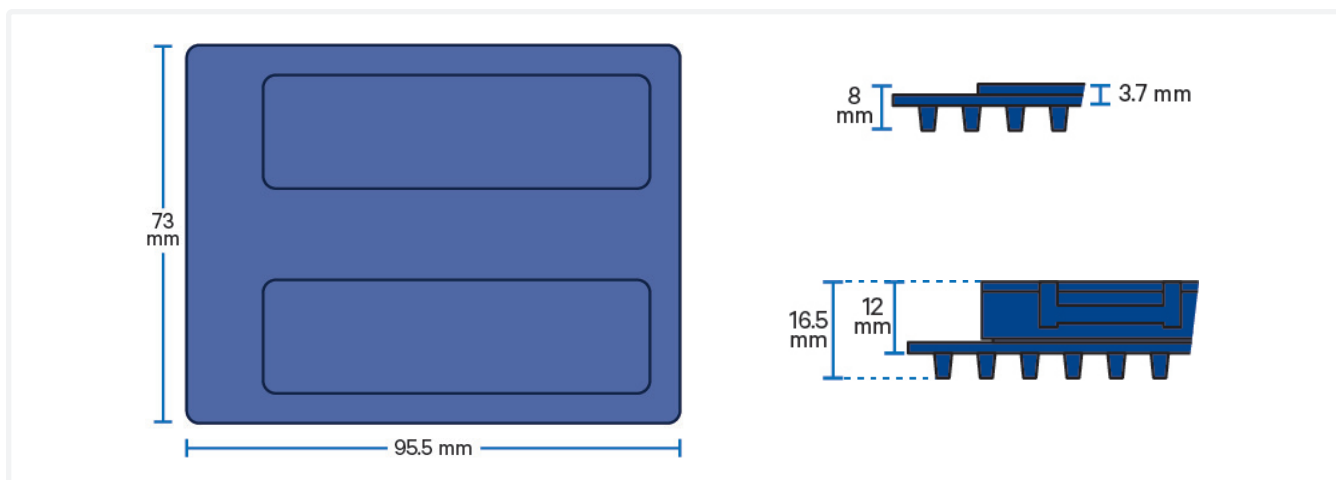
| Product | # | Kit and Part Number | Part Number (Item) |
|--------------------------|---|---|--------------------|
| Low Profile Plate Insert | 2 | Visium CytAssist Reagent Accessory Kit: 1000499 | 3000823 |
| 10x Magnetic Separator | 1 | | 230003 |

Recommended Thermal Cyclers

| Supplier | Description | Part Number |
|---------------------------------|--|---|
| Bio-Rad | C1000 Touch Thermal Cycler with 96-Deep Well Reaction Module | 1851197 |
| Eppendorf | MasterCycler Pro (discontinued) | North America 950030010 International 6321 000.019 |
| | MasterCycler X50s | North America 6311000010 |
| Thermo Fisher Scientific | Veriti 96-Well Thermal Cycler (discontinued) | 4375786 |

Thermal cycler must be able to accommodate the Low Profile Plate Insert (also referred to as the Low Profile Thermocycler Adapter):

- Well depth: 4.5 mm
- Distance between block and heated lid: 12 mm
- Reaction block dimensions 95.5 x 73 mm



Specific Reagents & Consumables

| Equipment For FFPE Tissue Sectioning & Section Placement | | | |
|--|---|-------------------|-------------|
| Item | Alternatives/Options | Vendor | Part Number |
| Microtome | Epredia HM 355S Automatic Microtome <i>Or any standard histology grade microtome</i> | Fisher Scientific | 23-900-672 |
| Microtome blade | Epredia MX35 Premier Disposable Microtome Blades, Low Profile | Fisher Scientific | 3052835 |
| Cool-Cut, <i>Optional</i> | Thermo Scientific Cool-Cut, <i>Optional</i> | Fisher Scientific | 77-112-0 |
| Section transfer system (STS) | Thermo Scientific Section Transfer System (STS), <i>Optional - If using Section Transfer System</i> | Fisher Scientific | 771200 |
| Probes | Fisherbrand Fine Precision Probe | Fisher Scientific | 12-000-153 |
| Forceps | Fisherbrand Curved Medium Point General Purpose Forceps | Fisher Scientific | 16-100-110 |
| Water bath | Premiere Tissue Floating Bath, Lighted <i>Or any equivalent water bath</i> | Fisher Scientific | 76278-820 |
| Section dryer oven | Epredia High Capacity Section Dryer | Fisher Scientific | A84600051 |
| Slide drying rack <i>If using section dryer oven</i> | StatLab TISSUE-TEK VERT 24 SLIDE RACK | Fisher Scientific | LWS2124 |
| Thermal cycler <i>Alternative to section dryer oven with Thermocycler Adapter</i> | C1000 Touch Thermal Cycler with 96-Deep Well Reaction Module | Bio-Rad | 1851197 |
| Brushes | Camel Hair Brushes <i>Or any equivalent paintbrush</i> | Ted Pella | 11859 |
| Green Marker, <i>Optional if marking area of interest</i> | Sharpie Argyle Green Permanent Marker | Sharpie | 1785396 |
| Blank Slides | Epredia Shandon ColorFrost Plus Slides | Fisher Scientific | 6776214 |
| | Fisherbrand Superfrost Plus Microscope Slides | Fisher Scientific | 12-550-15 |
| | Poly-Prep Slides | Millipore Sigma | P0425 |
| | Superfrost Slides | VWR | 48311-703 |
| Additional Materials | | | |
| Razor blades | | - | - |
| Ice bucket | | - | - |
| Dry ice | | - | - |
| Ultrapure/Milli-Q Water for Water Bath, <i>from Milli-Q Integral Ultrapure Water System or equivalent</i> | | - | - |

| For RNA Quality Assessment | | | |
|---|---|--------------------------|-------------|
| Xylene | Xylene, Reagent Grade | Millipore Sigma | 214736 |
| | Xylene, Histological Grade | Millipore Sigma | 534056 |
| Ethanol | Ethyl Alcohol, 200 Proof | Millipore Sigma | E7023 |
| | Ethanol absolute ≥99.5%, TechniSolv, pure (Europe Only) | VWR | 83813.360DP |
| RNA extraction | RNeasy FFPE Kit (50) | Qiagen | 73504 |
| Deparaffinization solution | Deparaffinization Solution | Qiagen | 19093 |
| RNase decontamination solution | RNaseZap RNase Decontamination Solution | Thermo Fisher Scientific | AM9780 |
| Nuclease-free water | Nuclease-free Water (not DEPC-Treated) | Thermo Fisher Scientific | AM9937 |
| Staining dishes <i>If working with hardset coverslipped slides. Alternative to Coplin jars.</i> | Staining Dish, White | VWR | 25608-906 |
| Coplin jars <i>If working with hardset coverslipped slides. Alternative to staining dishes.</i> | Glass Coplin Staining Jar, Screw Cap | VWR | 2100500-232 |
| Metal Block <i>If working with hardset coverslipped slides</i> | Light Labs 96-WELL ALUMINUM BLOCK | Fisher Scientific | NC9820161 |
| 1.5-ml centrifuge tubes | DNA LoBind Tubes, 1.5 ml | Eppendorf | 022431021 |
| 0.2 ml PCR 8-tube strips <i>Choose either Eppendorf, USA Scientific, or Thermo Fisher Scientific PCR 8-tube strips</i> | PCR Tubes 0.2 ml 8-tube strips | Eppendorf | 951010022 |
| | TempAssure PCR 8-tube strip | USA Scientific | 1402-4700 |
| | MicroAmp 8-TubeStrip, 0.2 ml | Thermo Fisher Scientific | N8020580 |
| | MicroAmp 8-cap Strip, clear | Thermo Fisher Scientific | N8010535 |
| Nanodrop/Qubit fluorometer | Nanodrop 2000c Spectrophotometers <i>Or any equivalent Nanodrop Alternative to Qubit Fluorometer</i> | Thermo Fisher Scientific | ND-2000C |
| | Qubit RNA BR Assay Kit | Thermo Fisher Scientific | Q10210 |
| | Qubit Assay Tubes | Thermo Fisher Scientific | Q32856 |
| | Qubit 4 Fluorometer | Thermo Fisher Scientific | Q33238 |
| Bioanalyzer/Tapestation <i>Choose Bioanalyzer or TapeStation based on availability & preference.</i> | RNA 6000 Pico Kit | Agilent | 5067-1513 |
| | 2100 Bioanalyzer Laptop Bundle | Agilent | G2953CA |
| | 4200 TapeStation | Agilent | G2991AA |
| | High Sensitivity RNA ScreenTape | Agilent | 5067-5579 |
| | High Sensitivity RNA ScreenTape Ladder | Agilent | 5067-5581 |
| | High Sensitivity RNA ScreenTape Sample Buffer | Agilent | 5067-5580 |

Tips & Best Practices



Tips & Best Practices section includes additional guidance



Signifies critical step requiring accurate execution



Troubleshooting section includes additional guidance

Sample Preparation

- Store FFPE tissue blocks at 4°C and avoid exposure to direct light to ensure even chilling and to preserve RNA integrity.

RNA Quality Assessment

- Assess RNA quality of the tissue block or archived sections before sectioning by calculating the percentage of total RNA fragments >200 nucleotides (DV200) of extracted RNA.
- Different factors may influence DV200 scores, such as specific tissue types, diseased or necrotic tissues, sample preparation, handling, loading concentration errors, or errors with ladder.

Section Thickness

- Recommended section thickness is 3–10 µm. Though this range of thicknesses was tested internally, most sections were cut at 5 µm, the thickness referenced throughout this protocol.

Water Bath Temperature & Section Floating Time

- Optimal water bath temperature and section floating time are critical for tissue section expansion. 42°C is the recommended water bath temperature for most tissues. However, water bath temperature may need optimization based on the tissue type.
- If the tissue is taking too long to expand, increase water bath temperature by 1–2 degrees and increase floating time.
- If the tissue is expanding too quickly and dissociating, decrease water bath temperature by 1–2 degrees and shorten floating time.
- Clean water bath according to manufacturer recommendations, followed by an RNase decontamination solution.

Section Placement on Blank Slides

- After section placement, blank slides are referred to as tissue slides.
- Before section placement, draw an outline of the allowable area on the back of the blank slide to ensure downstream compatibility with the Visium CytAssist instrument (Refer to [Determining Allowable Area](#)).
- If working with multiple sections on multiple tissue slides, place sections in the same location on the tissue slides for improved imaging efficiency.
- If placing multiple sections on a blank slide, ensure that the paraffin and/or tissue do not overlap.
- If multiple sections are placed on a blank slide, only one section from each tissue slide should be used for each Visium CytAssist Spatial Gene Expression Slide Capture Area.

Practice Section Placement

- Practice correct section placement using nonexperimental blocks.

Section Attachment

- Tissue block and section quality can affect the section attachment to blank slides.
- Carefully inspect the tissue block to gauge the extent of dehydration. Allow sufficient time in the ice bath to ensure proper hydration.
- Use a new, clean blade for sectioning each tissue type. Inspect the blade after every 20-25 sections and adjust to the blade areas that are not nicked or rough. Replace the blade after ~50 sections.
- Perform sectioning in a continuous motion to get a ribbon of sections. The sections should be separated during floating in the water bath. If floating multiple sections, monitor float time carefully.
- Ensure collected sections have the same thickness throughout experiments and replicates.
- Allow the section to float in the water bath until it is free of folds and wrinkles. Folds are associated with poor UMI capture and can be identified via H&E staining, by eye, or under the microscope before staining.
- Tissue detachment may occur due to factors such as:
 - Paraffin quality used during the tissue embedding process
 - Tissue block age
 - Tissue section thickness
 - Length of time used to infiltrate a tissue in paraffin.Refer to [Troubleshooting](#) for more information.



Optional Area of Interest Selection for Large Tissue Sections

If a tissue section does not fit completely within the Visium CytAssist Spatial Gene Expression Slide Capture Area (6.5 mm or 11 mm), a smaller area of interest should be defined. Refer to Figure A in the [Determining Allowable Area](#) section or the Visium CytAssist for Accessory Kit Instruction Quick Reference Card (CG000548).

Large Tissue Section in Paraffin Block



The area of interest should be:

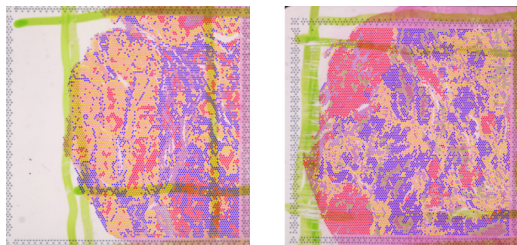
- Small enough to fit inside a well in the Visium CytAssist Tissue Slide Cassette gasket (refer to [Determining Allowable Area](#), Figure B for allowable area sizing). Tissue outside of the gasket will not be processed during the CytAssist workflow.
- Centered within the appropriate Capture Area size on the Tissue Slide Stage on the Visium CytAssist instrument

Define an area of interest after section placement by examining the tissue under a microscope. Area of interest can also be defined after H&E staining and imaging.

Annotate the area of interest on the back of the unstained tissue slide with a permanent marker to assist in Tissue Slide Cassette gasket alignment.

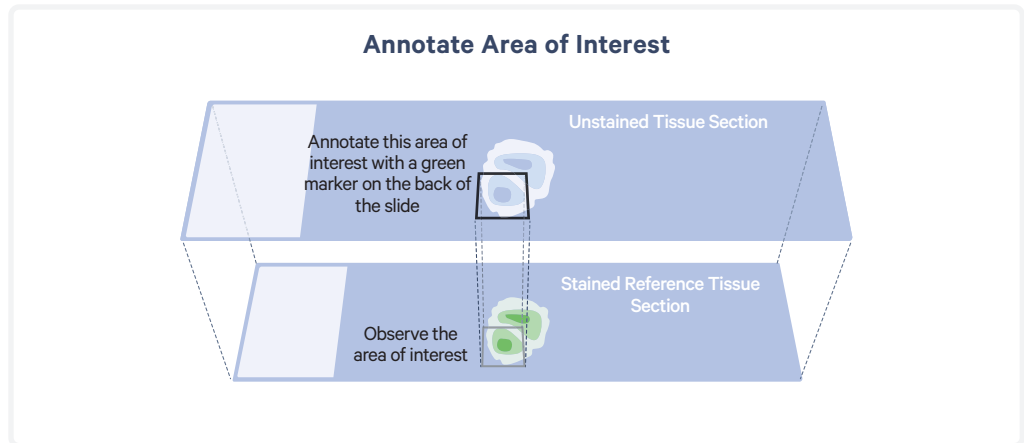
- Remove annotations before loading the tissue slide onto the instrument. Any remaining markings may cause automatic tissue registration to fail, resulting in a need for manual registration.
- If leaving a mark is desired, a green colored marker has the least impact on the automatic tissue registration process. 10x Genomics recommends using the Sharpie Argyle Green Permanent Marker (PN-1785396), or a marker with a similar hue, for minimal impact on automatic tissue registration.

Areas of Interest Annotated with Green Marker



If immunofluorescence staining is needed to define an area of interest, perform the following:

- a. Place a tissue section onto a blank slide as described in this Demonstrated Protocol.
- b. Stain the entire tissue section using any preferred method.
- c. Image the tissue.
- d. Annotate the area of interest on the back of the slide.
- e. Place a serial section onto a blank slide as described in this Demonstrated Protocol.
- f. Place the section onto the slide such that the area of interest is in the Allowable Area (refer to [Determining Allowable Area](#) for more information).
- g. Place the reference slide generated in step c face down on a flat, clean surface.
- h. Hover the unstained tissue slide upside down over the reference slide without allowing the slides to touch. See image below.



- i. After deparaffinization, annotate the area of interest on the back of the unstained tissue slide with a permanent marker to assist in Tissue Slide Cassette gasket alignment. 10x Genomics recommends using the Sharpie Argyle Green Permanent Marker (PN-1785396), or a marker with a similar hue, for minimal impact on automatic tissue registration.

Annotations may need to be readded before loading the tissue slide onto the instrument after the Eosin staining step in the main assay. If necessary, perform these steps again to reannotate the slide, or use the Tissue Slide Cassette gasket to draw annotations before disassembly.

- j. Repeat for any additional unstained tissue slides.
- k. Proceed with staining.

Determining Allowable Area

Use the following diagrams to verify that freshly placed tissue sections are compatible. Reference the images below to draw the allowable area on the back of blank slides (remove after tissue placement). Images are to scale if scaling settings are not modified. To verify, ensure that the first block in section A measures 6.5 x 6.5 mm.

A Ensure that the area of interest will fit within the Capture Area of a Visium CytAssist Spatial Gene Expression Slide.

Tissue sections larger than these Capture Areas may be placed on the slide, but only the tissue within the Capture Area will be processed by the CytAssist instrument.

Overlay the slide, centering the tissue on either square.

6.5 mm
Capture Area



11 mm
Capture Area

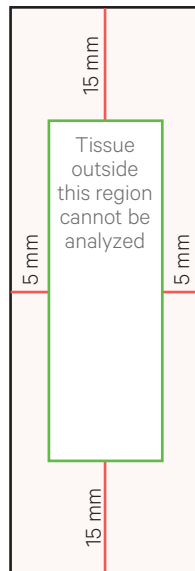


B If slide has no frosted areas, overlay on this diagram.

Tissue should lie within the green allowable area:

15 mm from top and bottom edges

5 mm from the sides

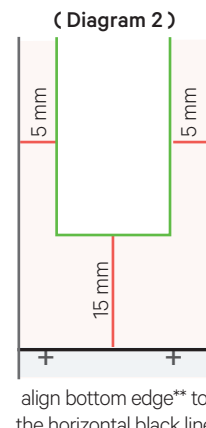
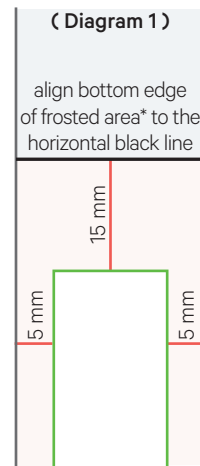


If slide has a frosted end and/or marks, overlay on diagram 1 and then on diagram 2.

Check the allowable area from both the top (1) and bottom (2) to ensure the tissue lies within the green allowable area (area is variable due to variability in the dimensions of frosted areas across slide brands):

15 mm from edge of frosted area/marks

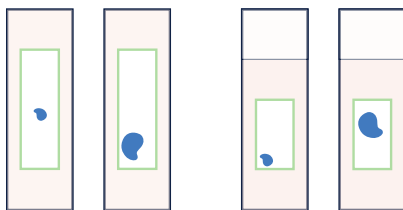
5 mm from the sides



*If text is present below frosted area of slide, align bottom of the text to the line

**If markings are present at bottom edge of slide, align markings to the + signs

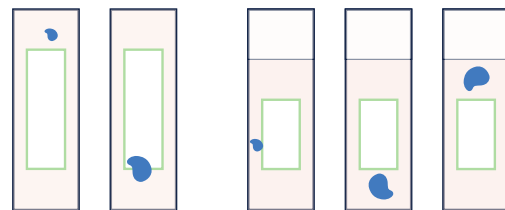
C Examples of Good Tissue Placement



Unfrosted Slide

Frosted Slide

Examples of Bad Tissue Placement



Unfrosted Slide

Frosted Slide

Visium CytAssist Tested Slides

The following slides have been tested for use with the Visium CytAssist Tissue Slide Cassette and instrument.

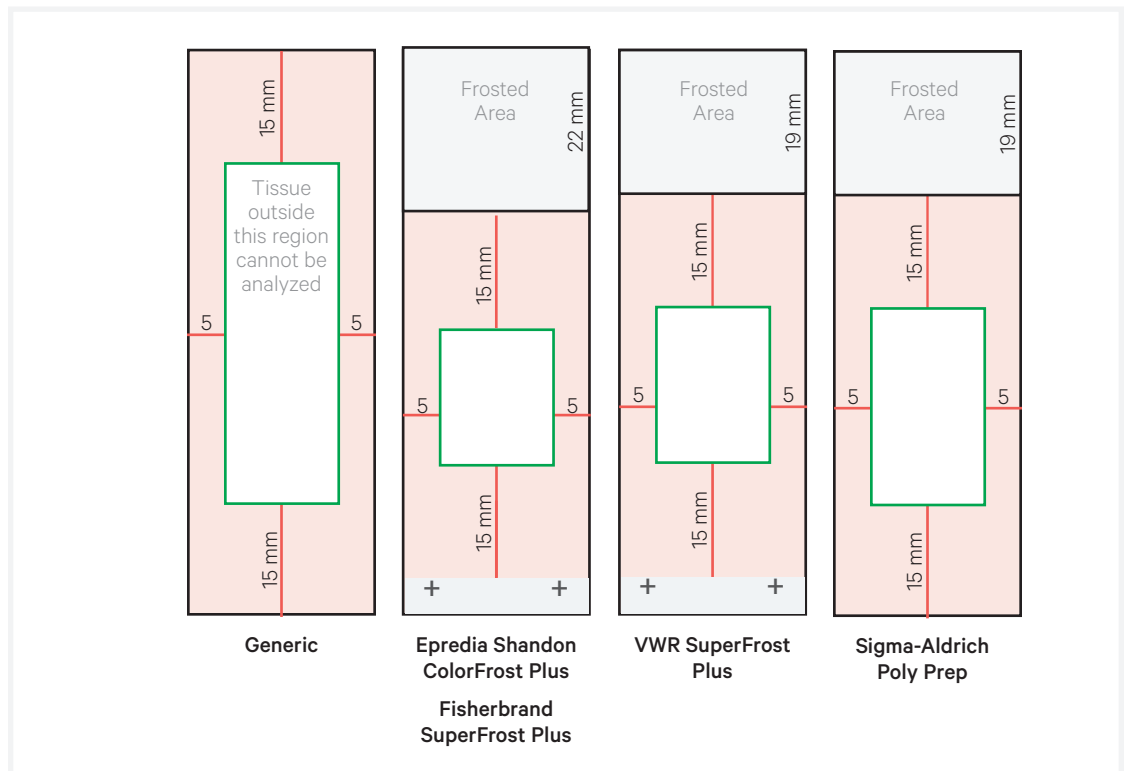
| Item | Length (mm) | Width (mm) | Thickness (mm) |
|---|-------------|------------|----------------|
| Epredia Shandon ColorFrost Plus Slides | 75.0 | 25.0 | 1.0 |
| Fisherbrand SuperFrost Plus Microscope Slides | 75.0 | 25.0 | 1.0 |
| Sigma-Aldrich Poly Prep Slides | 75.0 | 25.0 | 1.0 |
| VWR SuperFrost Plus Micro Slide, Premium | 75.0 | 25.0 | 1.0 |

If unsure of slide part number, refer to "generic" slide diagram below for general guidance (images not to scale). Diagrams for verifying that tissue sections are placed in the allowable area can also be found in the Visium CytAssist Quick Reference Cards - Accessory Kit (Document CG000548). The diagrams demonstrate allowable areas that are far enough away from frosted sections to not interfere with gasket closure during the CytAssist assay. Frosted sections include the opaque area of the slide as well as any etching on the slide.

While slides are specified as being 25 mm x 75 mm, manufacturing tolerances may lead to dimensions that are too small or large to be compatible with 10x Genomics products. Tissue slide dimensions must be within 24.8 mm - 25.3 mm in width and 74.4 mm - 76.2 mm in length to fit the Visium CytAssist Tissue Slide Cassette.

Minimum slide dimensions: 24.8 x 74.4 mm

Maximum slide dimensions: 25.3 x 76.2 mm



Handling Tissue Slides

- When immersing slides in reagent, ensure all tissue sections are immersed.
- Maintain tissue slides in a low moisture environment such as a desiccator, avoid exposure to direct light, and keep at room temperature.
- Tissue slides incubated at 42°C for 3 h and dried overnight at room temperature in a desiccator can be stored for up to two weeks at room temperature in a desiccator.

Tissue Slide Incubation (choose one method)

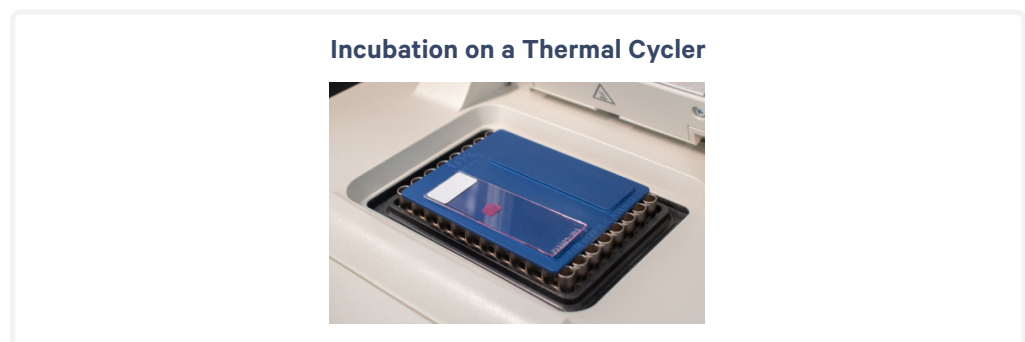
A. Incubation using a Section Dryer Oven:

- Place tissue slides in a slide drying rack on its side to prevent melted paraffin from disturbing adjacent tissue sections (if applicable).
- Close the lid when incubating the tissue slide in the oven.



B. Incubation using a Thermal Cycler:

- Position a Low Profile Thermocycler Adapter on a thermal cycler that is set at the incubation temperature. Low Profile Thermocycler Adapter must come to temperature before placing tissue slide.
- Ensure that the Low Profile Thermocycler Adapter is in contact with the thermal cycler surface uniformly.
- When incubating a tissue slide, position the tissue slide on the Low Profile Thermocycler Adapter with the side with tissue facing up.
- Ensure that the entire bottom surface of the tissue slide is in contact with Low Profile Thermocycler Adapter.
- DO NOT close the thermal cycler lid when incubating the tissue slide.
- Allow Low Profile Thermocycler Adapter to cool before removing from thermal cycler.



1. Hardset Coverslip Removal and RNA Quality Assessment for H&E Stained Archived Slides

Overview

This chapter provides guidance on hardset coverslip removal (coverslips that have been sealed with a reagent such as CytoSeal) and assessing RNA quality for archived slides.

Archived slides are defined as glass slides containing FFPE tissue sections that were placed in the past, where mounting of a coverslip was performed with CytoSeal or nail polish (hardset). Archived slides should have been imaged and stored at room temperature or 4°C. Over time, archived slides may experience RNA degradation; thus, freshly placed FFPE tissue sections are preferred for the Visium CytAssist assay.

These protocols should be performed on archived slides derived from the same tissue block as the slide that will be used for the full CytAssist assay. If this is not possible, an unimportant area of the section for processing can be scraped to assess RNA quality.

Before removal of hardset coverslip from the slide, verify that images of archived slides meet the specifications required for analysis as specified in Document CG000521. If images are absent or incompatible, before hardset coverslip removal, image the archived slide according to guidelines described in the Visium CytAssist Imaging Guidelines Technical Note (CG000521).



After hardset coverslip removal, H&E imaged and stained sections proceed directly to Destaining in the Visium CytAssist Spatial Gene and Protein Expression - Deparaffinization, H&E Staining, Imaging & Decrosslinking for FFPE Tissues Demonstrated Protocol (Document CG000658).

Remove Hardset Coverslip

Archived slides are dipped in xylene for a few minutes followed by rapid freezing to remove coverslips without damaging tissue sections.

Scrape Tissue Sample

Once hardset coverslip is removed, a small portion from a proximal section or an unimportant area of tissue from the section of interest is scraped to assess RNA quality.

Assess RNA Quality

RNA quality of the tissue on the archived slide is assessed by calculating the DV200 score of extracted RNA.

Mean RNA fragment size is a reliable determinant of RNA quality. This requires a measurement of the percentage of total RNA fragments >200 nucleotides (DV200) for RNA quality assessment upstream of library preparation. For more information on DV200, refer to [Appendix 1: DV200 Performance and Recommendations](#) and [Appendix 2: Example DV200 Traces](#).

Before starting RNA extraction, apply RNaseZap to a paper towel and wipe work surface, centrifuge rotor and shafts of the pipettes. Rinse with RNase-free water and then wipe dry. The electrodes of the Bioanalyzer can also be cleaned with RNaseZap by following Agilent decontamination procedure.

Proceed to Document CG000658

After assessing RNA quality, proceed directly to Destaining (step 3.2) in the Visium CytAssist Spatial Gene and Protein Expression - Deparaffinization, H&E Staining, Imaging & Decrosslinking for FFPE Tissue Demonstrated Protocol (Document CG000658).

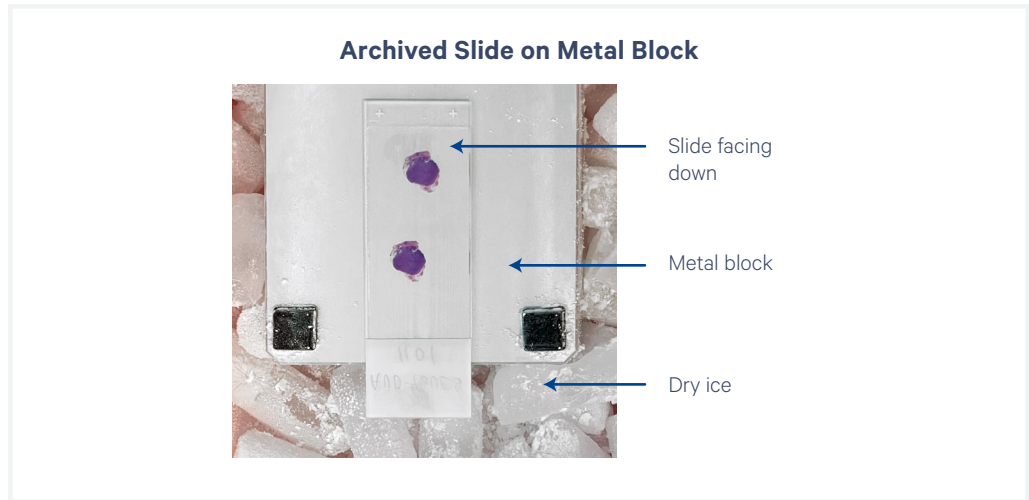
1.1 Preparation - Buffers

| Prepare fresh weekly. Process two slides per jar. | |
|---|--|
| Items | Preparation & Handling |
| <input type="checkbox"/> Xylene | Label two Coplin jars or staining dishes as Xylene Jar 1 and 2. Dispense 30 ml xylene in each. |
| <input type="checkbox"/> 100% Ethanol | Label two Coplin jars or staining dishes as 100% Ethanol Jar 1 and 2. Dispense 30 ml 100% ethanol in each. |
| <input type="checkbox"/> 96% Ethanol | Label two Coplin jars or staining dishes as 96% Ethanol Jar 1 and 2. Dispense 30 ml 96% ethanol in each. |
| <input type="checkbox"/> 70% Ethanol | Label one Coplin jar or staining dishes as 70% Ethanol Jar. Dispense 30 ml 70% ethanol. |
| Obtain | |
| Items | Preparation & Handling |
| <input type="checkbox"/> 100% Ethanol | |
| <input type="checkbox"/> Xylene | |
| <input type="checkbox"/> Nuclease-free water | |
| <input type="checkbox"/> Metal block | |
| <input type="checkbox"/> New Razor Blade | |
| <input type="checkbox"/> Dry Ice | |
| <input type="checkbox"/> 0.2 ml 8-tube Strip | |
| <input type="checkbox"/> RNeasy FFPE Kit | RNeasy MinElute Spin Column, Buffer PKD, Proteinase K, DNase Booster Buffer, DNase I Solution, RBC Buffer, and RPE Buffer are contained in this kit. |

1.2 Hardset Coverslip Removal

Xylene incubation steps should be performed in a fume hood. Either Coplin jars or staining dishes may be used. Before coverslip removal, image slide according to guidelines described in the Visium CytAssist Imaging Guidelines Technical Note (CG000521) and ensure they are compatible with the Space Ranger analysis pipeline. If archived slides have not been imaged, image per the above guidelines before removing the hardset coverslip.

- a. Cool a metal block on dry ice for **5–10 min.**
- b. Gently immerse archived slide in Xylene Jar 1. Secure the jar cap to prevent xylene loss.
- c. Incubate for **5 min.**
- d. Remove excess xylene from archived slide with a laboratory wipe and place on precooled metal block with the coverslipped tissue sections facing down.
- e. Wait **1 min.**



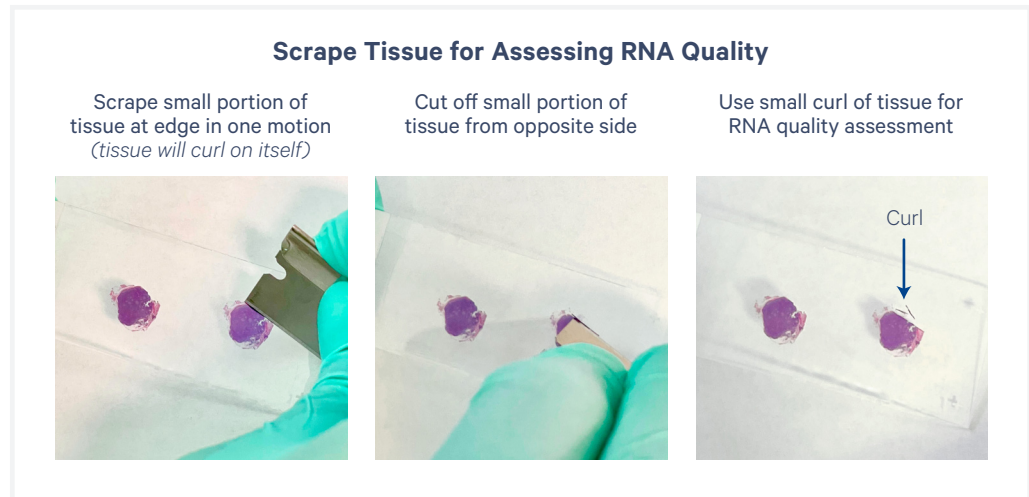
- f. Insert a clean blade a short distance between coverslip and archived slide on the shorter edge of the archived slide (see image below). Work slowly in small steps, keeping the archived slide on the cold metal block in between steps. Avoid touching tissue with the blade.



Exercise caution, as blade is sharp.



- g.** Immerse archived slide without coverslip 2x in Xylene Jar 1, then immerse for **10 min.**
- h.** Immerse archived slide 2x in Xylene Jar 2, then immerse for **10 min.**
- i.** Immerse archived slide 2x in 100% Ethanol Jar 1, then immerse for **3 min.**
- j.** Immerse archived slide 2x in 100% Ethanol Jar 2, then immerse for **3 min.**
- k.** Immerse archived slide 2x in 96% Ethanol Jar 1, then immerse for **3 min.**
- l.** Immerse archived slide 2x in 96% Ethanol Jar 2, then immerse for **3 min.**
- m.** Immerse archived slide 2x in 70% Ethanol Jar 1, then immerse for **3 min.**
- n.** Remove excess ethanol from archived slide carefully with a laboratory wipe. Do not touch tissue.
- o.** Select a small portion of the section that can be scraped for RNA quality assessment. Sections for RNA quality assessment should have a minimum size of 2 x 2 mm and minimum thickness of 5 µm. Practice scraping sections from test tissues.
- p.** Using a clean blade, scrape the small portion for RNA quality assessment in one motion, resulting in one curl.



- q.** Lift the curl using the blade and use a clean pipette tip to transfer curl to one 0.2-ml tube in a tube strip on ice. Store at **-80°C** for long-term storage or proceed immediately to RNA Extraction. If processing multiple archived slides, curls can be kept in tube strips on ice until ready for RNA extraction.
- r.** The remaining section on the archived slide may be used for the Visium CytAssist workflow or stored in a sealed slide mailer in a desiccator kept in the dark at **4°C** for up to **one week**.

1.3 RNA Extraction

- a. Prepare the thermal cycler with the following incubation protocol and start the program.

| Lid Temperature | Reaction Volume | Run Time |
|-----------------|-----------------|----------|
| 80°C | 160 µl | 30 min |
| Step | Temperature | Time |
| Pre-equilibrate | 56°C | Hold |
| Incubation 1 | 56°C | 00:15:00 |
| Incubation 2 | 80°C | 00:15:00 |

- b. Add **150 µl** of Buffer PKD to sample tube and pipette mix.
- c. Add **10 µl** of Proteinase K to sample tube and pipette mix.
- d. Add sample tube to thermal cycler and skip pre-equilibrate step.
- e. Pipette mix every **5 min** during Incubation 1 and 2 steps. Pipette mix without removing tube from block to prevent burns.
- f. Incubate a 2-ml microcentrifuge tube on ice for **3 min**.
- g. Transfer sample to the precooled 2-ml microcentrifuge tube after Incubation 2.
- h. Add **16 µl** of DNase Booster Buffer to tube.
- i. Add **10 µl** of DNase I Solution to tube. Pipette mix.
- j. Incubate at room temperature for **15 min**.
- k. Add **320 µl** of RBC Buffer to tube and pipette mix.
- l. Add **720 µl** of 100% Ethanol to tube and pipette mix.
- m. Transfer sample to RNeasy MinElute column. Do not allow sample to overflow in the column.
- n. Centrifuge column for **15 sec** at **8,000 rcf**.
- o. Repeat steps m-n until all sample has passed through the column.
- p. Add **500 µl** of RPE Buffer to column.
- q. Centrifuge column for **2 min** at **8,000 rcf**.
- r. Transfer column to a new 2-ml microcentrifuge tube.
- s. Centrifuge column for **5 min** at **maximum speed** with column lid open.
- t. Transfer column to a new 2-ml microcentrifuge tube.
- u. Add **12 µl** of nuclease-free water to column.
- v. Centrifuge column for **1 min** at maximum speed.
- w. Place RNA on ice or store at **-80°C** until ready to assess RNA quality.

- x.** Use **1 μ l** of sample for DV200 evaluation using either Agilent RNA 6000 Pico Kit or TapeStation (High Sensitivity RNA ScreenTape, RNA ScreenTape Sampler Buffer, RNA ScreenTape Ladder). Follow manufacturer's instructions for DV200 evaluation.
 - For more information on DV200, refer to [Appendix 1: DV200 Performance and Recommendations](#) and [Appendix 2: Example DV200 Traces](#).

After DV200 evaluation, proceed directly to Destaining (step 3.2) in the Visium CytAssist Spatial Gene and Protein Expression - Deparaffinization, H&E Staining, Imaging & Decrosslinking for FFPE Demonstrated Protocol (Document CG000658).

2. FFPE Tissue Sectioning & Section Placement

Overview

This chapter provides guidance on sectioning FFPE tissue blocks using a microtome and section placement on blank slides using a water bath. A Section Transfer System (STS) can also be used for section placement.

Exposing the Tissue

FFPE tissue block is placed in a microtome and cut to expose the tissue or face the block.

Sectioning

A section is taken from the block for RNA quality assessment.

RNA Quality Assessment

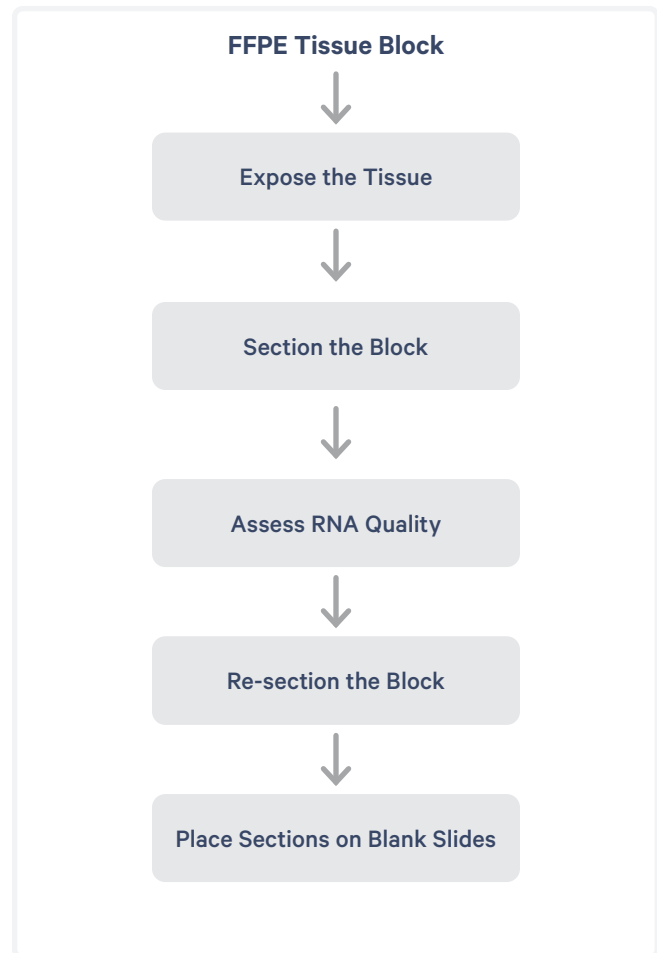
RNA quality of the tissue is assessed by calculating DV200 of RNA extracted from freshly collected tissue sections.

Re-sectioning

The tissue block is then sectioned by a microtome to generate sections for the blank slides.

Section Placement

Sections are collected in a water bath and are allowed to float on the water surface until they are flat and free from wrinkles and folds. The optimal floating time depends upon the specific tissue type and should be empirically determined when working with new tissue types. Expanded sections are then placed on blank slides.

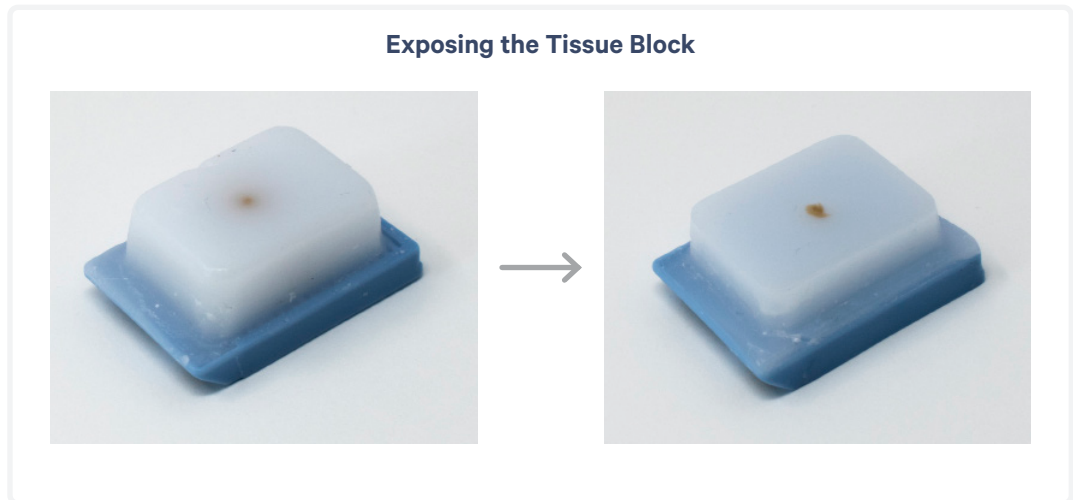


Practice section placement using a nonexperimental block before proceeding with tissue slides for the Visium workflow. Each tissue slide must only be used for only one CytAssist experimental run. DO NOT rerun tissue slides as this could affect assay performance.

2.1 Exposing the Tissue or Facing the Block

Before starting, wipe down all the surfaces and work areas with RNaseZap RNase decontaminating solution.

- a. Remove tissue blocks from storage. For a tissue block with already exposed tissue, proceed directly to RNA Quality Assessment (step 2.2).
- b. Set the microtome to the 15 μm setting.
- c. Place tissue block on the specimen clamp.
- d. Cut the tissue block at 15 μm until all of the edges of the tissue are exposed or until the area of interest is exposed. The block should be at **room temperature** during cutting.



2.2 RNA Quality Assessment of FFPE Tissue Block

This section provides guidance on determining the RNA quality of the tissue block by calculating DV200 of RNA extracted from freshly collected tissue sections.

Mean RNA fragment size is a reliable determinant of RNA quality. This requires a measurement of the percentage of total RNA fragments >200 nucleotides (DV200) for RNA quality assessment upstream of library preparation. For more information on DV200, refer to [Appendix 1: DV200 Performance and Recommendations](#) and [Appendix 2: Example DV200 Traces](#).

Before starting RNA extraction, apply RNaseZap to a paper towel and wipe work surface, centrifuge rotor and shafts of the pipettes. Rinse with RNase-free water and then wipe dry. The electrodes of the Bioanalyzer can also be cleaned with RNaseZap by following Agilent decontamination procedure.

- a. Set the microtome to the 10 μm setting and collect tissue sections of 10 μm thickness for RNA extraction. Discard the first few sections if the block was already exposed. The number of sections needed depends upon the tissue size. Refer to the RNA extraction kit manufacturer instructions to determine the appropriate number of sections. See below for guidance:
 - Collect ~4 sections for smaller tissues ($\leq 6.5 \times 6.5 \text{ mm}$)
 - Collect 1-2 sections for larger tissues ($\geq 6.5 \times 6.5 \text{ mm}$)
- b. Place the sections inside a **precooled** microcentrifuge tube. Sections may be stored at **-80°C** for **long-term** storage. For sections stored at **-80°C**, equilibrate to **room temperature** for **5 min** before adding the deparaffinization solution.
- c. Proceed to RNA extraction using RNeasy FFPE Kit and follow manufacturer's instructions.
- d. Using a Nanodrop or Qubit Fluorometer, measure RNA concentration to determine the appropriate dilution for DV200 evaluation using the RNA 6000 Pico Kit. RNA outside of the recommended concentration may lead to inaccurate DV200 evaluation.
- e. Store purified RNA at **-80°C** for **long-term** storage or **immediately** proceed to DV200 evaluation using either the Agilent RNA 6000 Pico Kit or TapeStation High Sensitivity Kit. Follow manufacturer's instructions (Agilent) for DV200 evaluation.

2.3 Sectioning

Section Collection:

- a. Place blocks in the ice bath, ensuring that the tissue part is fully submerged.
- b. Incubate on the ice bath for **10–30 min**. The incubation time depends upon the tissue type and the extent of dehydration.



Monitor the exposed tissue every 5–10 min during the ice bath incubation. The tissue surface should be smooth and shiny and free of bumps at the end of the incubation. For more information on tissue hydration, see Troubleshooting section.

FFPE Tissue Block in Ice Bath



- c. Carefully wipe off the excess oils from a 35X Ultra disposable blade using a laboratory wipe with 100% ethanol. Let the ethanol evaporate before proceeding. Always use new blades for sectioning each tissue type.



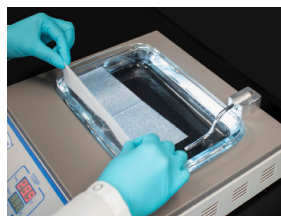
- d. Secure blade in disposable blade holder of the microtome and place knife guard over the blade to minimize injury. Ensure that the clearance angle is set to 10°.
- e. After hydration is complete, place the tissue block in the specimen clamp and align it with the blade.

- f. Fill a water bath with Milli-Q or ultrapure water. Water should be at **42°C** and free from bubbles & particulates. Remove bubbles & particulates, glide a laboratory wipe over the water surface. Repeat this step between sectioning if necessary.



42°C is the recommended water bath temperature for most tissues. However, water bath temperature may need optimization based on the tissue type. Determine optimal water bath conditions before tissue placement by practicing section placement on a blank slide with nonexperimental blocks. See Tips & Best Practices for guidance on optimizing water bath temperature. To better visualize the tissue sections, a Lighted Tissue Floating Bath with LED illumination can be used instead of standard water bath.

Remove Bubbles



- g.** Set the microtome to 5 μm for tissue sectioning and begin sectioning. For tissue blocks with exposed tissue, discard the first few sections and start collection on the subsequent sections.
- h.** To collect sections, place the paintbrush tip slightly above and parallel to the blade. Lift the section by lightly touching the edge with the paintbrush while rotating the wheel handle.
- i.** With the help of the brush, pick the section up. Immediately place section on the water bath surface, ensuring that the brush tip goes underneath and away from the section.
- j.** Proceed directly to Section Placement.

2.4 Section Placement

- a. Before proceeding with tissue slides intended for the Visium CytAssist workflow, practice section placement using nonexperimental blocks. Consider the following:
 - If placing multiple sections on the blank slide, ensure that sections do not overlap. Only one tissue section can be analyzed per tissue slide.
 - If placing a large section, refer to [Optional Area of Interest Selection for Large Tissue Sections](#) for information on selecting an area of interest.

Trace allowable area onto back of the blank slide with a laboratory marker before section placement to ensure compatibility. Refer to [Section Placement on Blank Slides](#) and [Visium CytAssist Validated Slides](#) sections for more information. Markings will need to be removed before alignment on the Visium CytAssist instrument.

- b. Allow sections to float for the time previously determined to be optimal.
- c. Hold the top of the blank slide and insert into the water, aligning the allowable area with the surface of the water while keeping the blank slide straight.
- d. Using the paintbrush or the probe, maneuver the section to the allowable area.




If sections float away from the blank slide, the blank slide can also be dipped into the water bath before sections are placed in the water.



- e. Pull the blank slide up and out of the water, ensuring there are no air bubbles trapped underneath. Set aside in a standing rack.
- f. Place the tissue slides in a slide drying rack in a section dryer and incubate for **3 h** in an oven at **42°C**. Alternatively, a thermal cycler set at **42°C** can be used for drying.

TIPS

See Tips & Best Practices for guidance on slide incubation.

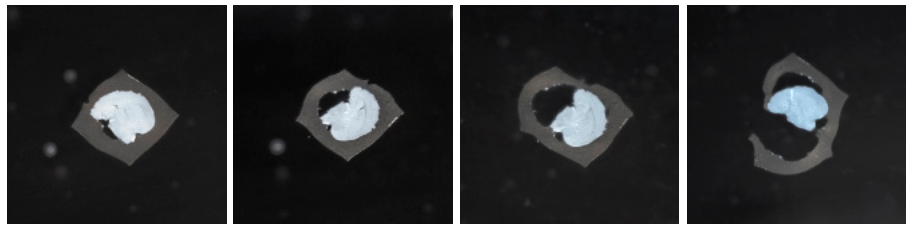
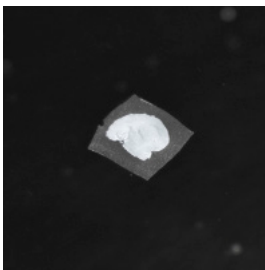
- g. Place in a desiccator and keep overnight at **room temperature** to ensure proper drying.
-  h. After overnight drying, proceed to deparaffinization and staining protocols (see References) or store the tissue slide containing dry tissue sections at **room temperature** in a desiccator for up to **2 weeks**.

Troubleshooting

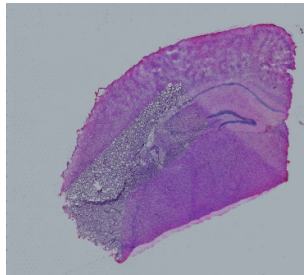
Ideal Floating Time Determination

Ideal Floating Time

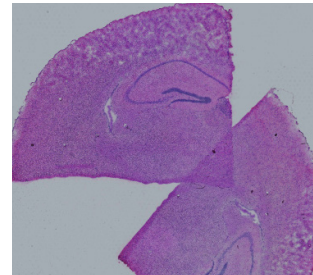
Section disintegration due to increased floating time



Incorrect Placement of Tissue Sections



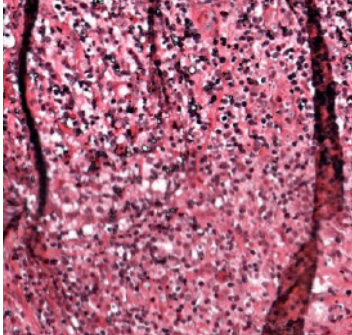
Folded tissue section



Overlapping sections

Common Artifacts that cause Detachment or Misleading Spatial Data

Wrinkles



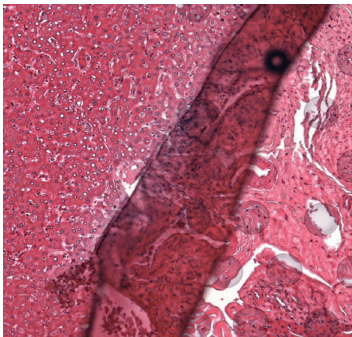
Causes

- Section compression (due to warm block or dull blade) during sectioning leads to wrinkle formation. These wrinkles become permanent when placed in the water bath.
- Accumulated wax or static electricity on microtome parts also contribute to section compression.
- Incorrect blade/clearance angle may cause compression.

Troubleshooting

- Ensure that the block is well hydrated.
- Adjust temperature down and increase float time.
- Gently and gradually lay FFPE sections onto water bath surface, lengthwise.
- Use a new blade.
- Ensure microtome is cleaned with 100% ethanol to minimize static and section compression (bunching on blade).
- Ensure blade/clearance angle is correct before sectioning.

Folds



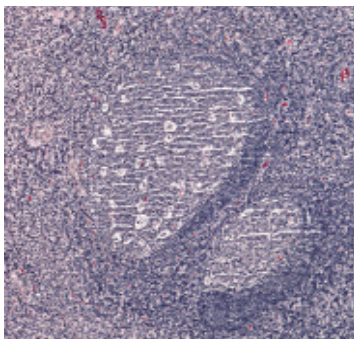
Causes

- Mostly happens when placing the section on the water bath, especially when the section is wavy.
- If the fold is at the edge, this most likely happened during sectioning or mounting on the slide.

Troubleshooting

- Gently and gradually lay FFPE ribbons or sections onto water bath surface, lengthwise.
- If sections curl during sectioning, gently flatten them with a brush before floating.

Venetian Blinds or Shatter



Causes

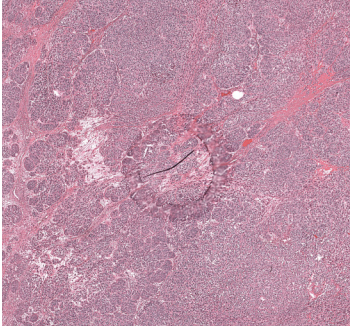
- Parallel lines in the section mostly appear due to dry tissue because of under-hydration of the block in the ice bath.
- Less likely due to dull blade or loose parts of the microtome.

Troubleshooting

- Increase incubation time of the block in ice bath.
- Tighten down components of microtome and make sure the blade is at a correct angle.

Common Artifacts that cause Detachment or Misleading Spatial Data

Air Bubbles



Causes

- Air bubbles from the bottom of the water bath can rise and stick under the section.

Troubleshooting

- Using a brush, gently remove the bubbles from the bottom of the water bath before floating the sections.

Waves



May be block-specific and easily observed under the microscope right after the section is picked up from the water bath. Minor waves can disappear after longer flotation in the water bath. Larger waves can create wrinkles or folds that are permanent after drying of the section.

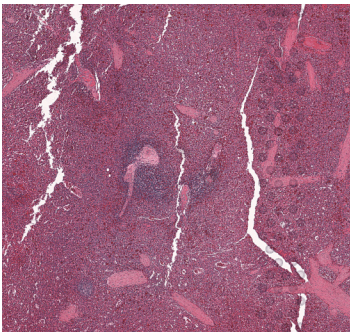
Causes

- Tissue incompletely infiltrated with wax absorbs water faster during hydration step. When sections from such blocks are floated, the parts that absorbed enough water will have difficulty flattening and become wavy.

Troubleshooting

- Chill the block on a cold block or on ice avoiding contact with water.
- Submerge the block for 5–15 min in the ice bath for gentle hydration.
- Increase flotation times and/or temperature of the water bath.

Cracks



Causes

- Dry and over-processed tissue can crack during sectioning.
- The cracks that are created before tissue embedding will be filled with wax when section is observed under the microscope after sectioning and wax will be washed away after deparaffinization or H&E staining.

Troubleshooting

- Prolonged hydration on the ice bath will most likely reduce the cracks.
- There is no solution for cracks created before tissue was embedded in wax.

Common Artifacts that cause Detachment or Misleading Spatial Data

Sweating



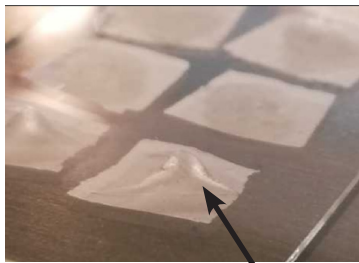
Causes

- Inadequate dehydration or under-processing of the tissue causes parts of the section to attract water and create this blistering effect. Such sections can disintegrate in the water bath.
- The blisters consist of:
 - Xylene or xylene substitutes if the cause is under-processing and insufficient removal of xylene or
 - Water droplets if the cause is inadequate dehydration.

Troubleshooting

- Be cautious about how long the block is kept in ice bath. Long incubation time in ice bath can impact section quality and thus should be avoided.
- Long flotation time in the water bath should be avoided.

Water Retention



Water retention under the section

Causes

- Sections from tissues that are under-processed or excessively hydrated before sectioning may retain water under the section mounted on a slide.
- Water will accumulate under section and cause uneven drying ultimately leading to detachment.

Troubleshooting

- Gently flicking the slide will help to get rid of the water. This is one of the most overlooked cause of detachment.

Disintegrating/Exploding Section



Causes

- Sections from tissues that are under-processed and not sufficiently infiltrated with wax can rapidly absorb water and explode or gradually disintegrate in the water bath.
- Tissues that are not sufficiently dehydrated can show similar phenotypes.
- Floating at high temperatures (42-50°C) can exacerbate the disintegration.

Troubleshooting

- Chilling of the block should be mostly performed on ice or cold block (30–60 min).
- Exposure to water in ice bath during chilling should be kept to a minimum (5–10 min).
- Flotation water bath temperature should be lowered to 38–40°C.

Appendix

A1: DV200 Performance and Recommendations

DV200 is a broad measurement of RNA quality and is influenced by factors including:

- Tissue block age, type and composition
- Region selected for RNA extraction
- Presence of diseased or necrotic regions
- Depth of section
- Fixation method
- Miscellaneous upstream tissue handling and processing

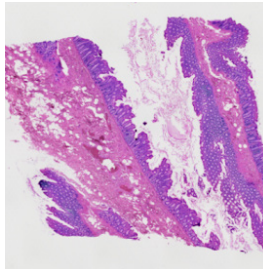
Assessment of multiple healthy and cancer tissue samples demonstrate a positive correlation between DV200 score and median UMI counts and panel genes detected with the Visium Spatial Gene Expression workflow at 20,000 raw reads per spot. Based on these observations, tissue samples with a DV200 of >30% are more likely to yield robust and reproducible results.

However, tissue samples with a low DV200 score have demonstrated compatibility with the assay with lower UMI and gene counts. For example, a DV200 of ~15% may yield ~6,000 UMI counts and ~3,800 panel genes detected at 20,000 raw reads per spot. Below are examples data with low DV200 score adjacent colon tissue sections from two different donors. In this example, the low DV200 score is likely due to the heterogeneous composition of the tissue, where ~70% of the section is comprised of low UMI count subcutaneous tissue.

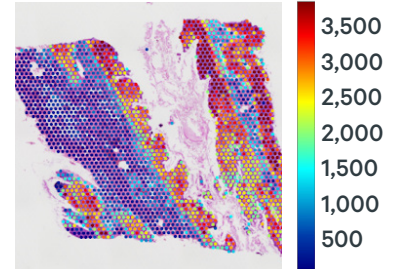
The following representative data were generated from freshly placed tissue sections with the Visium CytAssist Spatial Gene Expression assay.

Donor 1 - Adjacent Normal Colon, DV200 = 12.43

H&E Stain

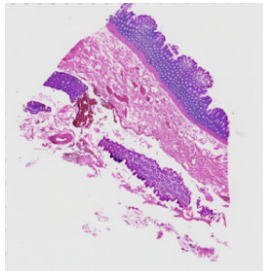


UMI Counts

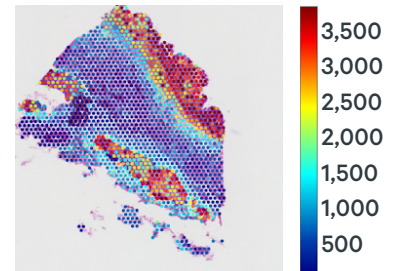


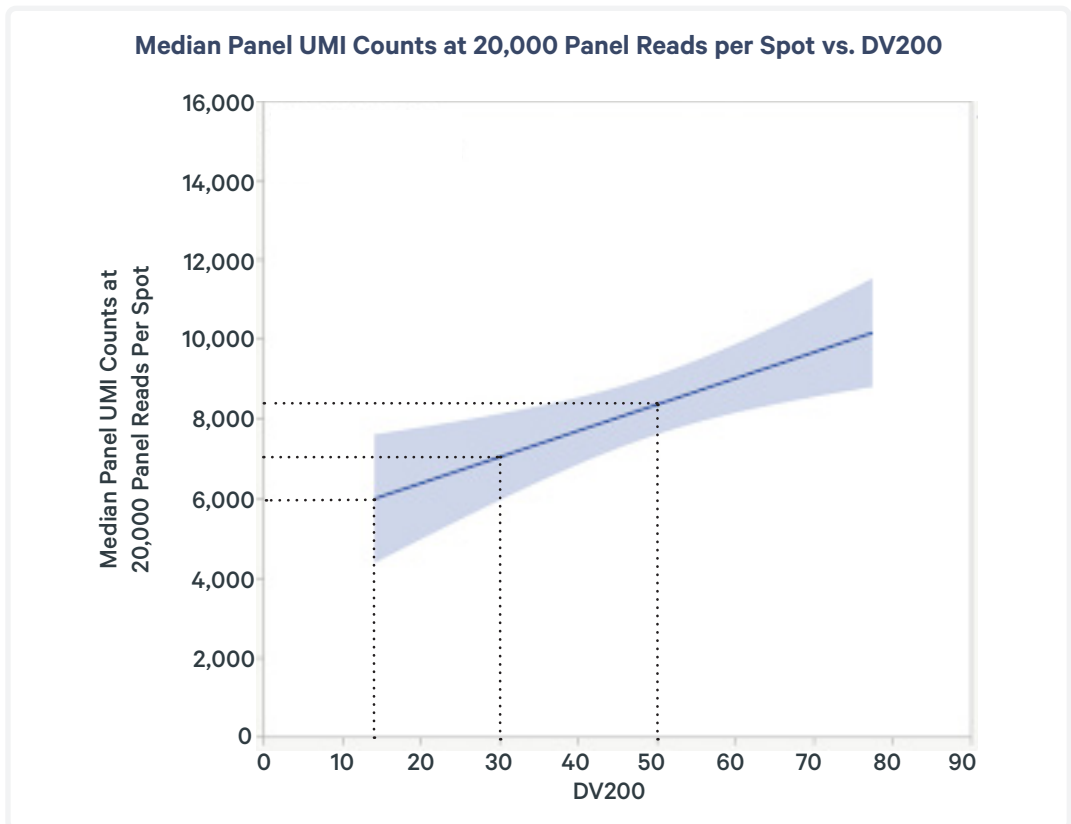
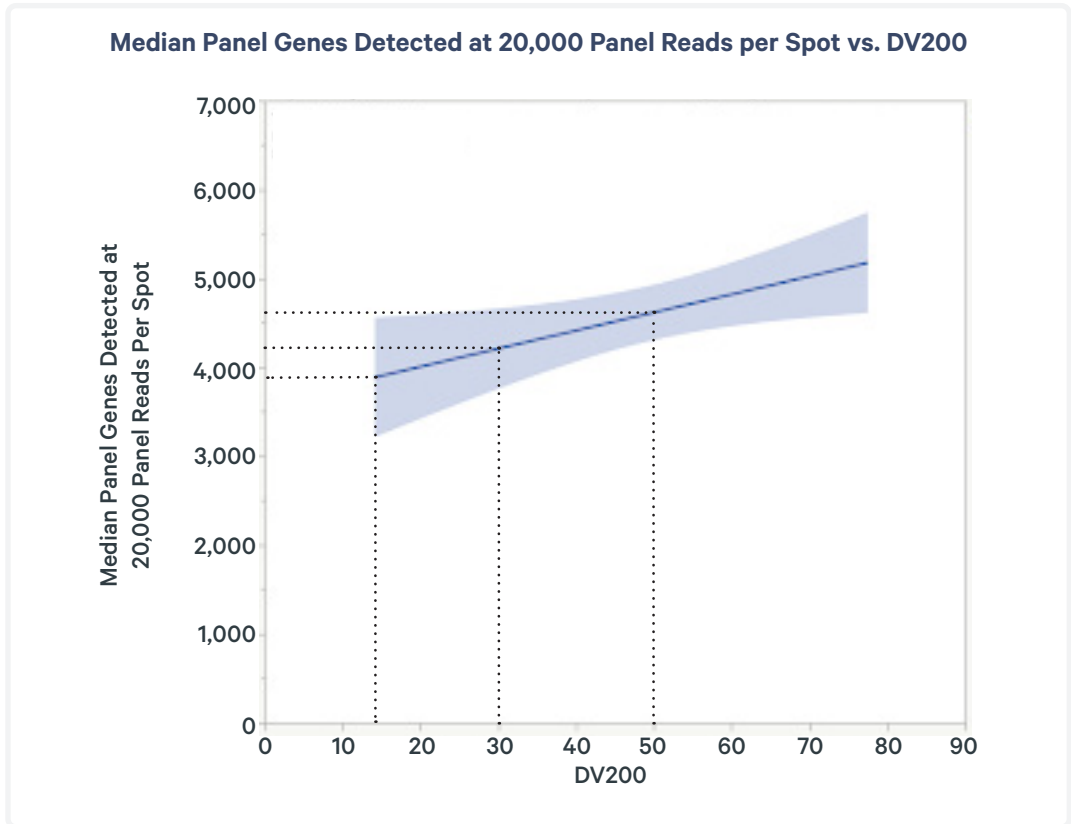
Donor 2 - Adjacent Normal Colon, DV200 = 25.78

H&E Stain



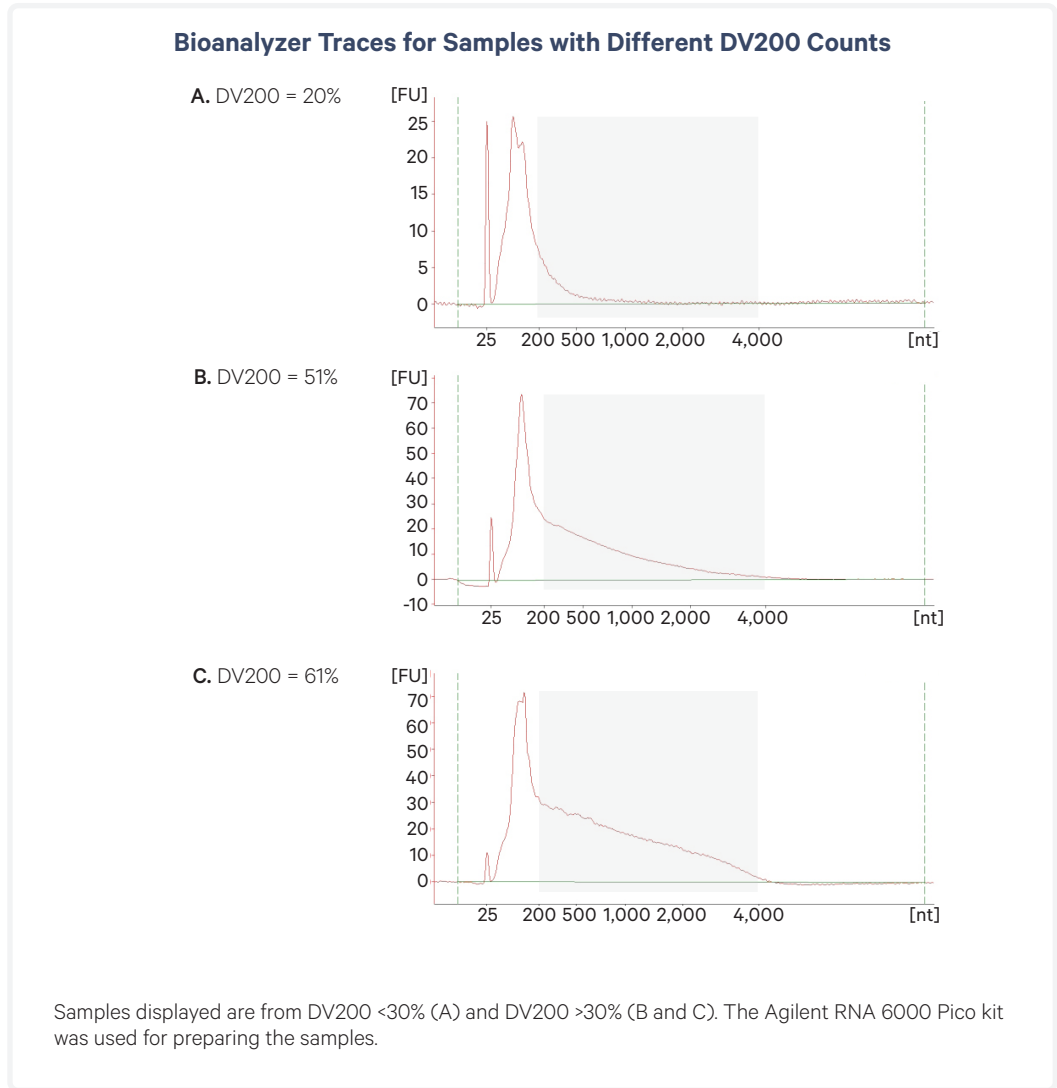
UMI Counts





A2: Example DV200 Traces

Representative Bioanalyzer traces of RNA extracted from different tissue types for which DV200 was calculated are shown below.



A3: Confirming Tissue Antigen Integrity

Before starting the Visium CytAssist Spatial Gene and Protein Expression assay, an immunofluorescence stain may be performed to confirm antigen integrity in the tissue section. Consult the Visium CytAssist Spatial Gene and Protein Expression – Deparaffinization, Decrosslinking, IF Staining & Imaging for FFPE Tissue (CG000659) for full instructions. Consult the 10x Genomics support website for a list of antibody clones used in 10x Genomics antibody panels.

References

- Visium CytAssist Spatial Gene and Protein Expression – Deparaffinization, H&E Staining, Imaging & Decrosslinking for FFPE Tissue (CG000658).
- Visium CytAssist Spatial Gene and Protein Expression – Deparaffinization, Decrosslinking, IF Staining & Imaging for FFPE Tissue (CG000659).
- Visium CytAssist for Accessory Kit Instruction Quick Reference Card (CG000548)
- Karigoudar, M, et al. Alternative Rapid Methods for Coverslip Removal: A Comparative Study. *Journal of Clinical and Diagnostic Research* 13:1-2, 2019

Document Revision Summary

| | |
|------------------------|--|
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| Title | Visium CytAssist Spatial Gene and Protein Expression – FFPE Tissue Preparation Guide |
| Revision | Rev A |
| Revision Date | May 2023 |

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