CG000256 Rev D

## **QUICK REFERENCE CARDS**

# **Chromium Connect**

#### **FOR USE WITH**

Chromium Connect, PN-1000171 Chromium Connect, PN-1000180



## **System Components**

- 1 Front Panel
- 2 Left side of the power base assembly:



- (a) Thermal Cycler Power Supply
- System Power SupplyOne or two independent circuits possible
- © Fuses/Fuse covers (two 12A fuses, two 4A fuses)
- 3 Right side of the power base assembly:
  - a USB port: external devices
  - (b) USB port: optional HEPA CAP hood
  - © USB port: Thermal Regulator
  - d Ethernet to customer Wired Network
  - Power plug: optional HEPA CAP hood
  - f Power plug: Thermal Regulator
- 4 Touchscreen
  USB port on monitor: User External Drive
  (download log files/upload setup files)
- (5) Gantry (liquid-handling device)
- 6 Barcode Scanner
- 7 Deck Area
- (8) Chromium Automated Controller
- Thermal Regulator
   (kept on/underneath benchtop)
  - Main power cable for Thermal Regulator (plugs into 3f)
  - **b** USB port
  - © ID port
  - d External Sensor port



REMOVE POWER FROM BOTH AC INLETS PRIOR TO ANY SERVICE ON THERMAL CYCLER OR SYSTEM.

Information also available in Chromium Connect Instrument User Guide (CG000180) and in the Software Menu Options.

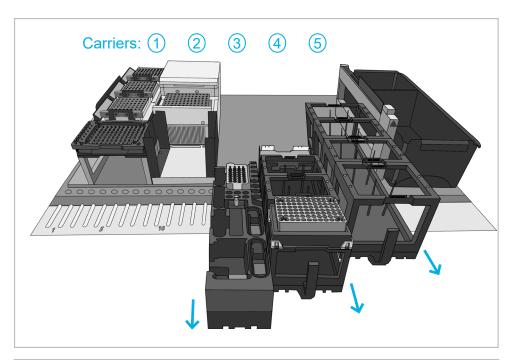


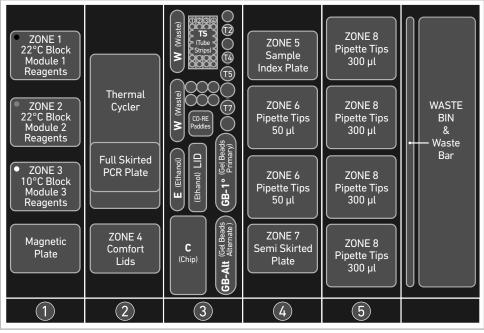


Chromium Next GEM Automated Single Cell Gene Expression 3' or 5' Assays

#### **Deck Area Layout**

The deck includes five carriers. Carriers 1-2 are stationary while Carriers 3-5 slide out for loading/unloading items. The items displayed in the layout and table are specific to the assays listed above.

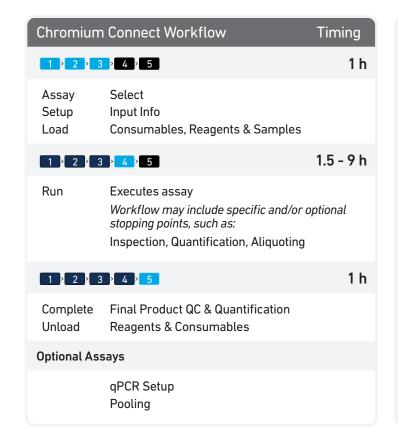


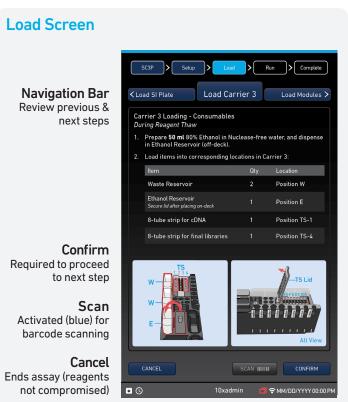


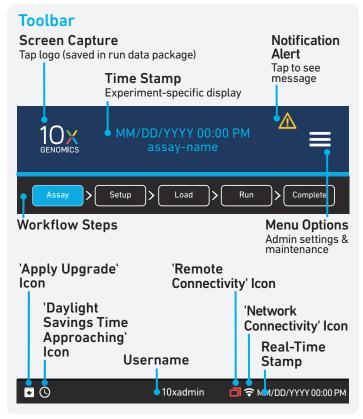
Zone	Item
Carrier 1	Stationary
ZONE 1	22°C Block, Reagent Strips
ZONE 2	22°C Block, Reagent Strips
ZONE 3	10°C Block, Reagent Strips
-	Magnetic Plate
Carrier 2	Stationary
-	Thermal Cycler
-	Full Skirted PCR Plate (within Thermal Cycler)
ZONE 4	ComfortLids
Carrier 3	Rails: 15-18, Lights: 4
W	Waste Reservoir
TS-1 to TS-4	Tube Strips
T2 T4 T5 T7	Dynabeads™ MyOne™ SILANE Primer Glycerol Partitioning Oil
-	CO-RE Paddles
E	Ethanol Reservoir
LID	Lid for Ethanol Reservoir
GB-1°	Gel Beads Primary
GB-Alt	Gel Beads Alternate
С	Chip
Carrier 4	Rails: 19-24, Lights: 6
ZONE 5	Sample Index Plate
ZONE 6	Pipette Tips 50 μl
ZONE 7	Semi Skirted Plate
Carrier 5	Rails: 25-30, Lights: 6

### Software Orientation

Run Screen







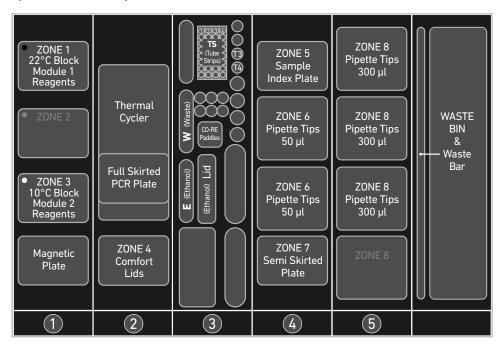




Chromium Next GEM Automated Single Cell 5' V(D)J Amplification Assay

#### **Deck Area Layout**

The deck includes five carriers. Carriers 1-2 are stationary while Carriers 3-5 slide out for loading/unloading items. The items displayed in the layout and table are specific to the assay listed above.



Chromium	Connect Workflow V(D)J Amplification	Timing
1 > 2 > 3	1 h	
Assay Setup Load	Select Input Info cDNA, Reagents & Consumables	
1 > 2 > 3	<mark>} 4 </mark>	7 h
Run	V(D)J Amplification 1 & 2 V(D)J Amplification QC Library Construction, Cleanup & Sample Indexing	
1 > 2 > 3	4 > 5	2 h
Complete Unload	Final Library QC & Quantification Reagents & Consumables	
Optional Assays		1.5 h
	qPCR Setup Pooling	

Zone	Item
Carrier 1	Stationary
ZONE 1	22°C Block, Reagents, Module 1
ZONE 2	-
ZONE 3	10°C Block, Reagents, Module 2
-	Magnetic Plate
Carrier 2	Stationary
-	Thermal Cycler
-	Full Skirted PCR Plate (within Thermal Cycler)
ZONE 4	ComfortLids
Carrier 3	Rails: 15-18, Lights: 4
W	Waste Reservoir
TS	Tube Strips
T3 T4	Primer Mix 2 Primer Mix 1
T4	Primer Mix 1
T4 CP	Primer Mix 1  CO-RE Paddles
T4 CP E	Primer Mix 1  CO-RE Paddles  Ethanol Reservoir  Lid for Ethanol Reservoir
T4 CP E LID	Primer Mix 1  CO-RE Paddles  Ethanol Reservoir  Lid for Ethanol Reservoir
T4 CP E LID Carrier 4	Primer Mix 1  CO-RE Paddles  Ethanol Reservoir  Lid for Ethanol Reservoir  Rails: 19-24, Lights: 6
T4 CP E LID Carrier 4 ZONE 5	Primer Mix 1  CO-RE Paddles  Ethanol Reservoir  Lid for Ethanol Reservoir  Rails: 19-24, Lights: 6  Sample Index Plate
T4 CP E LID Carrier 4 ZONE 5 ZONE 6	Primer Mix 1  CO-RE Paddles  Ethanol Reservoir  Lid for Ethanol Reservoir  Rails: 19-24, Lights: 6  Sample Index Plate  Pipette Tips 50 µl

## **Tips & Best Practices**

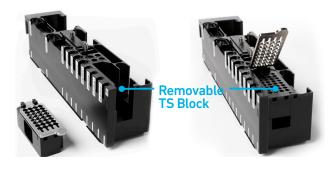
#### **Carrier Handling**

- Establish a clean space near the instrument for placing sliding carriers during loading.
- To scan barcodes, slide carrier in SLOWLY and follow software prompts (chime & flashing lights).
- Practice sliding the carriers completely off the deck and replacing them back using rails.



#### **Deck Loading - Consumables**

- Follow touchscreen for assay-specific handling.
- Ensure correct barcode orientation (on tubes and racks) as prompted by the touchscreen.
- The deck orients the A1 position to the back left corner of the instrument.
- Keep chip and gasket in sealed package until prompted to load.
- DO NOT use chips or gaskets specific to other 10x Genomics protocols.
- Optional removal of the TS block from Carrier 3 may facilitate loading tube strips.
- When loading PCR plates, press down firmly.
- When positioning the sample index plate into Carrier 4, the plate will audibly click into place and rear black clips will be visible.



#### **User Interaction**

- CSV file upload enabled for Assay Setup
- For sample prep, refer to assay-specific Automation Kit User Guide.
- The touchscreen provides a window for sample prep during which reagents are safely held on deck.
- Optional emulsion check requires user interaction ~30–50 min into the run.

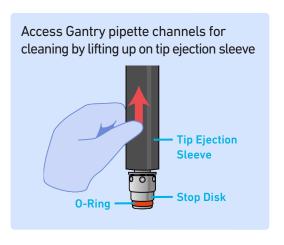
#### **Deck Loading - Reagents**

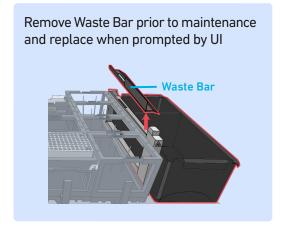
- Follow touchscreen for assay-specific handling.
- Ensure that no air gaps or precipitate remain at the bottoms of tubes.
- Prepare and dispense 80% ethanol off-deck to avoid spilling on consumables (e.g. chip).
- Carrier 3 gel bead positions allow use of one full tube strip or two partially used tube strips (adding up to 8 tubes total). Use GB-Primary first.
- Reagent thaw begins at specified points on touchscreen, with continued loading during thaw.
- Module loading: Use black handles to lift lids. Load from row 1 (back to front), inserting barcode end onto pin.
- Press black handles while closing lids.
   DO NOT allow lids to forcefully snap shut.
- DO NOT touch module mirrors. Smudges can prevent barcode scanning. Wipe clean when necessary.



## Maintenance

Running Maintenance via Software	Daily 15 min	Weekly 40 min
Inspect for Condensation, Dust & Smudges	✓	✓
Clean Touchscreen Lens-cleaning tissue and nuclease-free water	✓	✓
Empty Waste Bin & Liquid Waste	✓	✓
Channel Tests  Pipette Channel Tightness Test Capacitive Liquid Level Detection Test	✓	✓
Clean Deck, Rails, Carriers, Mirrors & Walls Wipe down with laboratory wipes to minimize sources of particles and fibers 70% isopropanol or microcide on metal Deionized water or microcide on nonmetal		<b>✓</b>
Clean Gantry Channels  Lens-cleaning tissue and nuclease-free water		✓
Calibration		✓
Chromium Automated Controller Self Check		✓





#### References

- Chromium Connect Instrument User Guide (Document CG000180)
- Chromium Connect Specifications (Document CG000255)
- 10x Genomics Support website

#### **Support**

Email: support@10xgenomics.com

10x Genomics 6230 Stoneridge Mall Road Pleasanton, CA 94588 USA

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Document Revision Summary

**Document Number** CG000256

Title Chromium Connect Quick Reference Cards

**Revision** Rev C to Rev D

**Revision Date** March 2023

#### **Specific Changes:**

• Deck Layout map now labels the positions that hold reagent tubes more universally on page 3.

• User Interface screens and workflow table updated on page 4.

• New best practice for loading sample index plate into carrier on page 6.

• Hardware change to gantry pipette channel illustration on page 7.

#### **General Changes:**

• Updated for general minor consistency of language and terms throughout.