10x GENOMICS

User Guide | CG000582 | Rev A

Xenium
In Situ Gene Expression

Probe Hybridization, Ligation & Amplification

For use with:

Xenium Slides & Sample Prep Reagents (2 slides, 2 rxns) PN-1000460

Xenium Decoding Consumables (1 run, 2 slides) PN-1000487

Xenium Mouse Brain Gene Expression Panel (2 rxns) PN-1000462

Xenium Human Breast Gene Expression Panel (2 rxns) PN-1000463

Xenium Custom Gene Expression Panel (up to 50 genes) PN-1000464

Xenium Custom Gene Expression Panel (51 to 100 genes) PN-1000561

Xenium Instrument Accessory Kit Module A PN-1000530

Notices

Document Number

CG000582 | Rev A

Legal Notices

Instrument & Licensed Software Updates Warranties

Updates to existing Instruments and Licensed Software may be required to enable customers to use new or existing products.

Support

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Document Revision Summary

Document Number
CG000582

Title
Xenium In Situ Gene Expression User Guide

Revision
Rev A

Revision Date
December 02, 2022

Specific Changes

General Changes

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Introduction

Reagent Kits

Xenium In Situ Gene Expression Reagent Kits

Refer to SDS for handling and disposal information.

Xenium Slides & Sample Prep Reagents - (2 slides, 2 rxns) PN-1000460

Xenium Slides & Sample Prep Reagents
(2 slides, 2 rxns), PN-1000460
Store at −20°C

		#	PN
	Xenium Probe Hybridization Buffer	1	2000390
	Xenium Post Hybridization Wash Buffer	1	2000395
	Xenium Ligation Buffer	1	2000391
	Xenium Ligation Enzyme A	1	2000397
	Xenium Ligation Enzyme B	1	2000398
	Xenium Amplification Mix	1	2000392
	Xenium Amplification Enzyme	1	2000399
	Reducing Agent B	1	2000087
	Xenium Autofluorescence Mix	1	2000753
	Xenium FFPE Tissue Enhancer	1	2000798
	Xenium Nuclei Staining Buffer	1	2000762
	Perm Enzyme B	1	3000553
	Xenium Slides (2 pack)	1	3000941

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All items, except Xenium FFPE Tissue Enhancer (PN-2000798) and Perm Enzyme B (PN-3000553), are needed for this workflow.

Xenium Decoding Consumables - (1 run, 2 slides) PN-1000487

Xenium Decoding Consumables
(1 run, 2 slides), PN-1000487
Store at ambient temperature

	#	PN
Xenium Cassette Kit (2 cassettes + 16 lids)	1	1000566
Extraction Tip	1	2000757
Pipette Tips	1	3000866
Xenium Buffer Cap	1	3000949
Xenium Objective Wetting Consumable	1	2000749
Deionized Water	1	3001198
Xenium Sample Wash Buffer A	1	3001199
Xenium Sample Wash Buffer B	1	3001200
Xenium Probe Removal Buffer	1	3001201

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Only the Xenium Cassette Kit (2 cassettes + 16 lids) is needed for this workflow.

Xenium Mouse Brain Gene Expression Panel - (2 rxns) PN-1000462

Xenium Mouse Brain Gene Expression Panel
(2 rxns), PN-1000462
Store at −20°C

	#	PN
Xenium Probe Dilution Buffer	1	2000393
Xenium Mouse Brain Gene Expression Probes	1	2000825

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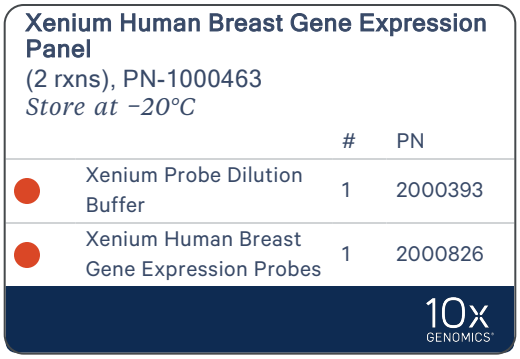
Xenium Human Breast Gene Expression Panel - (2 rxns) PN-1000463



Xenium Human Breast Gene Expression Panel
(2 rxns), PN-1000463
Store at −20°C

		#	PN
●	Xenium Probe Dilution Buffer	1	2000393
●	Xenium Human Breast Gene Expression Probes	1	2000826

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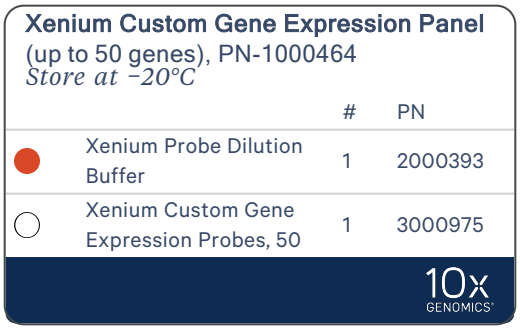
Xenium Custom Gene Expression Panel - (up to 50 genes) PN-1000464



Xenium Custom Gene Expression Panel
(up to 50 genes), PN-1000464
Store at −20°C

		#	PN
●	Xenium Probe Dilution Buffer	1	2000393
○	Xenium Custom Gene Expression Probes, 50	1	3000975

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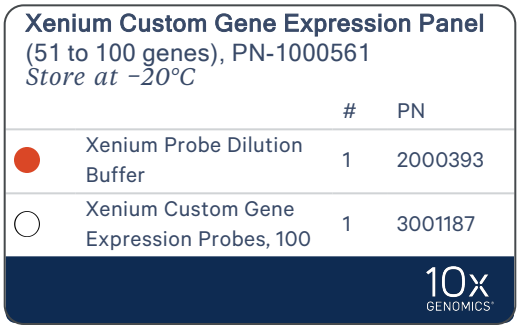
Xenium Custom Gene Expression Panel - (51 to 100 genes) PN-1000561



Xenium Custom Gene Expression Panel
(51 to 100 genes), PN-1000561
Store at −20°C

		#	PN
●	Xenium Probe Dilution Buffer	1	2000393
○	Xenium Custom Gene Expression Probes, 100	1	3001187

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Refer to the 10x Genomics website for the most updated list of available panels.

Xenium Instrument Accessory Kit Module A PN-1000530

Xenium Instrument Accessory Kit Module A
PN-1000530
Store at ambient temperature

	#	PN
Waste Bottle	1	3000955
Xenium Waste Tip Tray	1	3000957
Xenium Thermocycler Adaptor	1	3000954

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Only the Xenium Thermocycler Adaptor (PN-3000954) is needed for this workflow.

Recommended Thermal Cyclers

Supplier	Description	Part Number
Bio-Rad	C1000 Touch Thermal Cycler with 96-Deep Well Reaction Module	1851197

Thermal cycler requirements if reactions are performed on a slide:

- Temperature-controlled lid
- 96 deep-well block or 0.2 ml block configuration
- The thermal cycler must be able to accommodate the Xenium Thermocycler Adaptor:
  - Well depth: 4.5 mm
  - Distance between block and heated lid: 12 mm
  - Reaction block dimensions: 95.5 x 73 mm

Additional Kits, Reagents & Equipment

The listed items have been tested by 10x Genomics and perform optimally with the assay. **Substituting materials may adversely affect system performance.** For items with multiple options listed, choose option based on availability and preference. Refer to the manufacturer's website for regional part numbers.

Item	Description	Supplier	Part Number (US)
Plastics			
1.5 ml tubes	DNA LoBind Tubes, 1.5 ml	Eppendorf	022431021
	Low DNA Binding Tubes, 1.5 ml	Sarstedt	72.706.700
15 ml tubes	15 ml PP Centrifuge Tubes	Corning	430791
50 ml tubes	Self-Standing Polypropylene Centrifuge Tubes (50 ml), sterile	Corning	430921
Pipette tips	Tips LTS 20UL Filter RT-L20FLR	Rainin	30389226
	Tips LTS 200UL Filter RT-L200FLR	Rainin	30389240
	Tips LTS 1ML Filter RT-L1000FLR	Rainin	30389213
Kits & Reagents			
Nuclease-free Water	Nuclease-free Water (not DEPC treated)	Thermo Fisher Scientific	AM9937
TE Buffer	TE Buffer, TRIS-EDTA, 1X Solution, pH 8.0	Thermo Fisher Scientific	BP24731
PBS	PBS - Phosphate Buffered Saline (10X) pH 7.4, RNase-free	Thermo Fisher Scientific	AM9624
10% Tween-20	Tween 20 Surfact-Amps Detergent Solution (10% solution)	Thermo Fisher Scientific	28320
Ethanol	Ethyl alcohol, Pure (200 Proof, anhydrous)	Millipore Sigma	E7023-500ML
Equipment			
Pipettes	Pipet-Lite LTS Pipette L-20XLS+	Rainin	17014392
	Pipet-Lite LTS Pipette L-200XLS+	Rainin	17014391
	Pipet-Lite LTS Pipette L-1000XLS+	Rainin	17014382
Mini centrifuge	VWR Mini Centrifuge *(or any equivalent mini centrifuge)*	VWR	76269-064
Thermomixer	Eppendorf ThermoMixer C *(or any equivalent Thermomixer)*	Eppendorf	5382000023
Thermoblock	Eppendorf SmartBlock 1.5 mL	Eppendorf	5360000038

Item	Description	Supplier	Part Number (US)
	(or any equivalent Thermoblock)		
	Eppendorf SmartBlock 2.0 mL *(or any equivalent Thermoblock)*	Eppendorf	5362000035
Blank Slides	Shandon ColorFrost Plus Slides 25 x 75 x1 mm (Optional)	Thermo Fisher Scientific	6776214
	Fisherbrand Premier Plain Glass Microscope Slides (Optional)	Thermo Fisher Scientific	12-544-4
Additional Materials			
Waterbath			
Thermal Cycler *(C1000 Touch Thermal Cycler with 96-Deep Well Reaction Module, Bio-Rad, 1851197)*			
Ice bucket			
Vortex			
Ultrapure/Milli-Q Water for Water Bath (recommended), *from Milli-Q Integral Ultrapure Water System or equivalent*			

This list may not include some standard laboratory equipment.

Protocol Steps & Timing

Steps	Timing	Stop & Store
Day 1		
Step 1: Probe Hybridization		
1.1 Buffer Preparation	20 min	
1.2 Probe Hybridization	16-24 h (overnight)	
Day 2		
Step 2: Post Hybridization Wash		
2.1 Post Hybridization Wash	35 min	
Step 3: Ligation		
3.1 Ligation	~2 h	
Step 4: Amplification		
4.1 Amplification	~2 h	
4.2 Post Amplification Wash	15 min	STOP 4°C overnight or ≤4 days
Step 5: Autofluorescence Quenching		
5.1 Autofluorescence Quenching	45 min	STOP 4°C overnight or ≤4 days (in the dark)
5.2 Nuclei Staining	20 min	STOP 4°C overnight or ≤4 days (in the dark)

Stepwise Objectives

Xenium In Situ Gene Expression assays RNA at the subcellular level by using targeted probes in formalin fixed & paraffin embedded (FFPE) or fresh frozen (FF) tissue sections. FFPE tissue sections placed on Xenium Slides are deparaffinized and decrosslinked as described in Xenium In Situ for FFPE - Deparaffinization & Decrosslinking (Demonstrated Protocol – CG000580). FF tissue sections placed on Xenium slides are fixed and permeabilized as described in Xenium In Situ for Fresh Frozen - Fixation & Permeabilization (Demonstrated Protocol – CG000581).

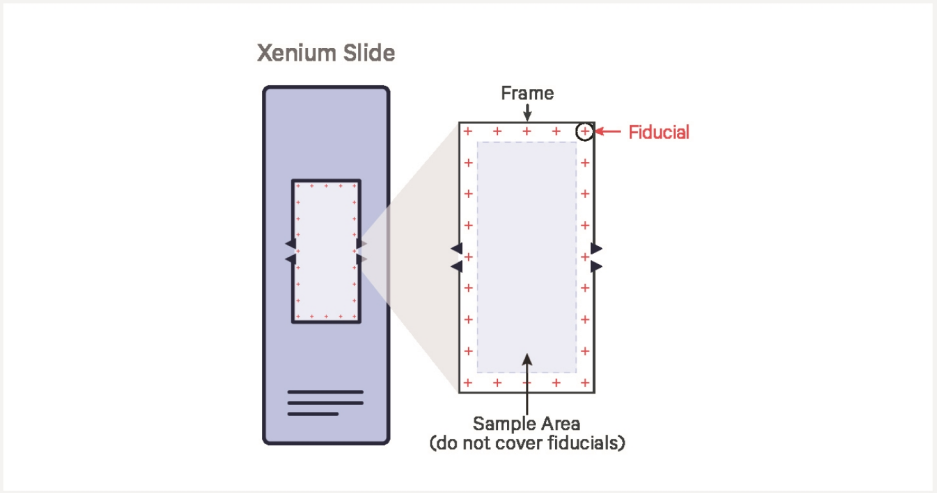
Pre-designed probe panels, with optional custom probe panels, are then added to the tissue. Each circularizable DNA probe contains two regions that hybridize to the target RNA and a third region that encodes a gene-specific barcode. The two ends of the probes bind the target RNA and are ligated to generate a circular DNA probe. Following ligation, the circularized probe is enzymatically amplified, generating multiple copies of the gene-specific barcode for each RNA target.

Xenium slides containing FFPE or FF tissue sections are then loaded for imaging and analysis on the Xenium Analyzer instrument for high-throughput, automated in situ analysis. Fluorescently-labeled oligos bind to the amplified DNA probes. Cyclical rounds of fluorescent probe hybridization, imaging, and removal generate optical signatures specific for each barcode, which are converted into a gene identity. Identified transcripts can be visualized using Xenium Explorer software.

This document outlines the protocol for generating Xenium In Situ Gene Expression data from FFPE and FF tissue sections placed on Sample Areas of a Xenium slide.

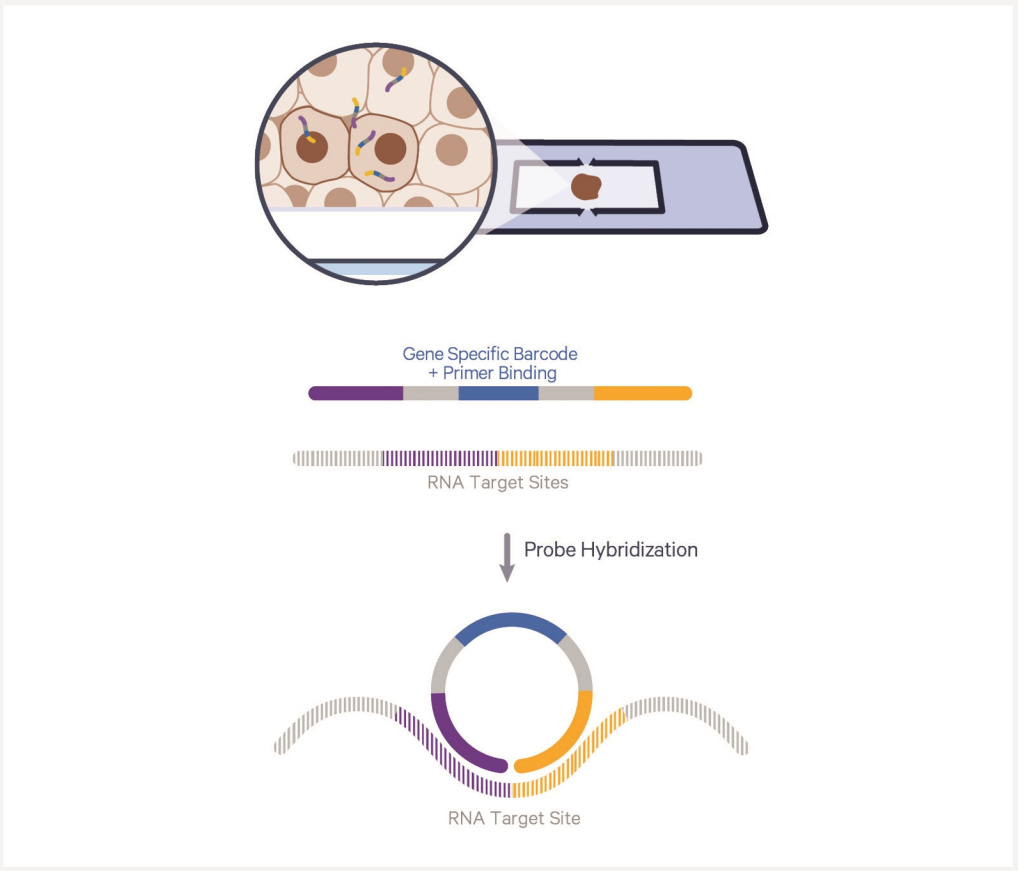
Xenium Slide

The Xenium slide has one Sample Area measuring 10.45 x 22.45 mm and is defined by a fiducial frame. The imageable area, measuring 12 mm x 24 mm, includes the area within the Sample Area + fiducial frame. FFPE or FF tissue sections are placed within the Sample Area for downstream processing and analysis.



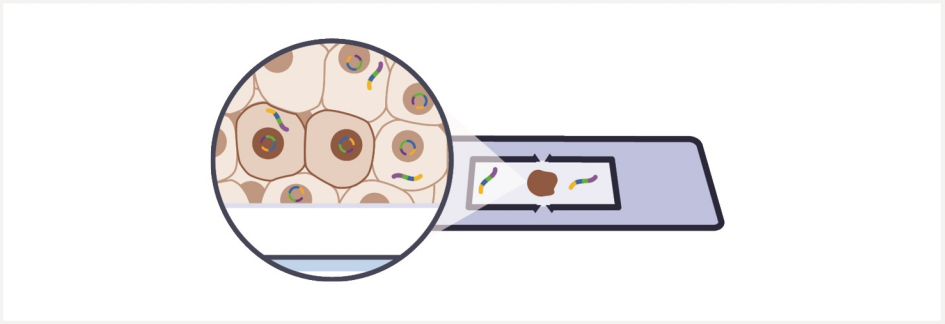
Step 1: Probe Hybridization

Pre-designed DNA probes, alone or in combination with custom DNA probes, are added to the FFPE or FF tissue sections. The DNA probes are flanked by two regions that independently hybridize to the target RNA and also contain a gene-specific barcode sequence. The probes hybridize to their complementary target RNA in an overnight incubation.



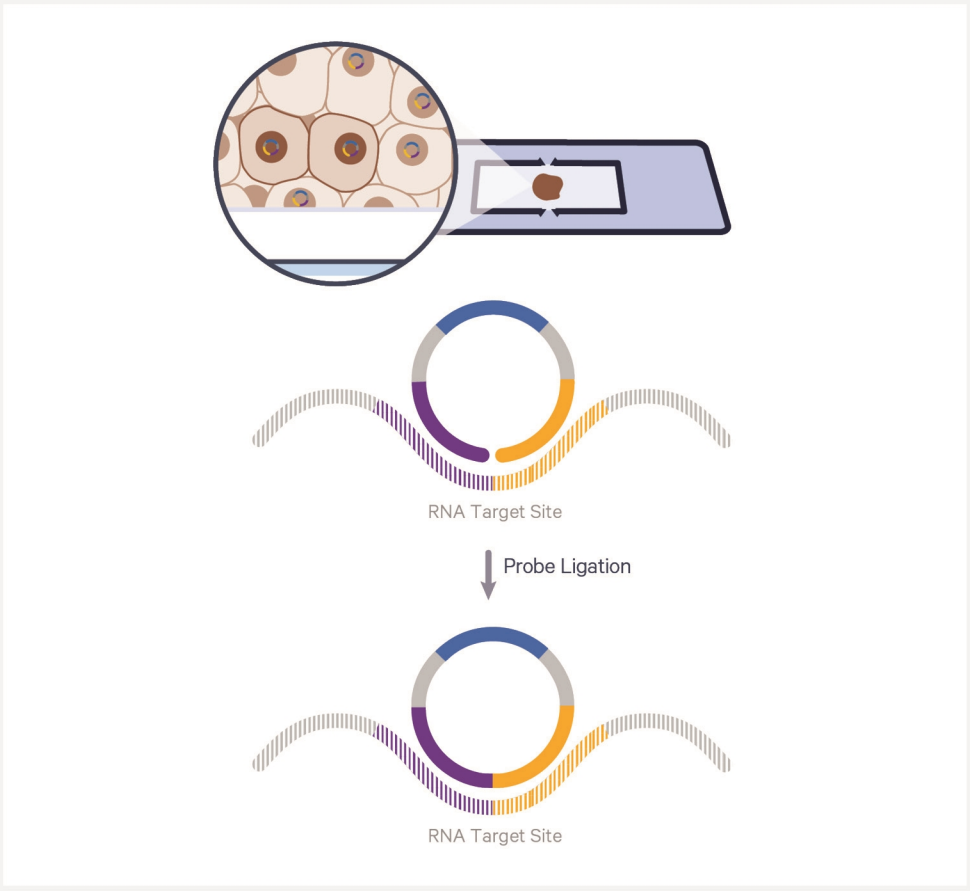
Step 2: Probe Hybridization Wash

Excess, unbound probes are washed away in the post hybridization wash step.



Step 3: Ligation

After removal of unbound probes, a ligase is added to seal the junction between the probe regions that have hybridized to RNA. Ligation of the probe ends on the targeted RNA region generates a circular DNA probe. This ligation ensures a unique level of probe specificity to the target region.

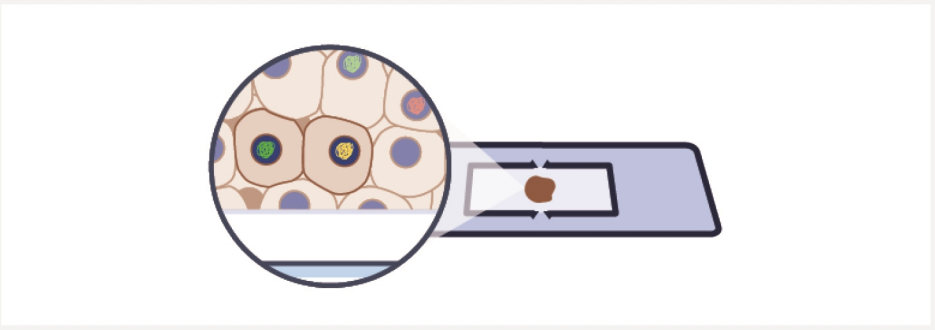


Step 4: Amplification

The ligation products are enzymatically amplified. Hundreds of copies of the gene specific barcode are generated during the amplification process.

Step 5: Autofluorescence Quenching

Autofluorescence Quenching diminishes unwanted autofluorescence and enhances signal-to-noise ratio in the treated FFPE and FF tissue sections. Next, nuclei are stained to assist in identification of tissue or regions of interest during an instrument overview scan. Finally, tissue sections on Xenium slides assembled into Xenium Cassettes are loaded into the Xenium Analyzer for imaging and decoding.



Tips & Best Practices

Icons

Tips & Best Practices section includes additional guidance

Signifies critical step requiring accurate execution

Troubleshooting section includes additional guidance

General Reagent Handling

- Fully thaw reagents at indicated temperatures. Thoroughly mix reagents before use.
- When pipette mixing reagents, unless otherwise specified, set pipette to 75% of total volume.
- Keep all enzymes and Master Mixes on ice during setup and use, unless otherwise stated.
- Promptly move reagents back to the recommended storage.

Pipette Calibration

- Follow manufacturer's calibration and maintenance schedules.

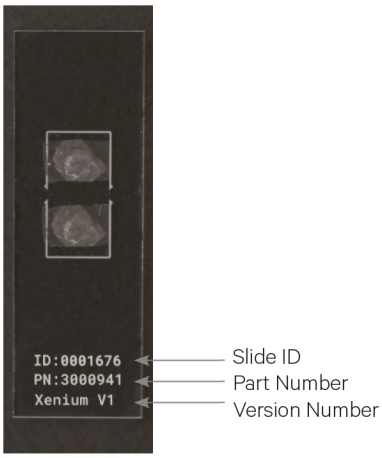
Custom Probe Handling

- Custom probe panels are optional and can be used in addition to pre-designed probe panels.
- Custom probes are delivered lyophilized and must be resupsended prior to use. See Probe Hybridization on page 34 for more details.
- Record the Custom Panel Design ID and Slide Number before starting the workflow. This information is critical for identifying the correct electronic decode file when setting up the Xenium Analyzer in downstream steps.

Xenium Slide Handling

- Always wear gloves when handling slides.
- The bottom of the slide is indicated by the etched label, which should be readable when in the proper position.
- The tissue sections should always be placed within the Sample Area on etched label side of the slide.
- Hold the slide on the label. DO NOT touch the tissue sections or near fiducials.
- Minimize exposure of the slides to sources of particles and fibers.
- Keep the slide cassette flat on the bench when adding reagents to the Sample Area.
- Ensure that no absorbent surface is in contact with the reagents on the slide during incubation.
- When pipetting reagent onto a slide, avoid generating bubbles. Avoid pipetting directly onto the tissue.

Xenium Slide with Tissue Sections



Processing a Single Xenium Slide

- Xenium reagent kits are sufficient for two reactions, and for optimal Xenium Analyzer throughput, two slides should be run at the same time.

- It is possible to perform the Xenium In Situ Gene Expression workflow with a single slide. To do this, ensure the following best practices are followed for optimal assay performance:
  - Assemble a mock Xenium Cassette using a blank slide and a cassette from the Xenium Cassette Kit (2 cassettes), PN-1000566.
  - Insert the blank slide into the Xenium Cassette. Cassettes should be assembled following the instructions in Troubleshooting for Xenium Cassette Assembly.
  - Attach a Xenium Cassette Lid from the Xenium Cassette Kit (2 cassettes), PN-1000566 to the cassette containing the blank slide following Tips & Best Practices for Xenium Lid Application. It is not necessary to add liquid to the slide well before adding the lid.
  - For all incubation steps with the thermal cycler lid closed, ensure the mock slide cassette is placed alongside the Xenium slide cassette containing tissue on the Thermocycler Adaptor.

Reagent Addition to Wells

- Place assembled cassette flat on a clean work surface.
- Dispense and remove reagents along the side of the well without touching the tissue sections and without introducing bubbles.
- Always cover the Sample Area completely when adding reagents to the well. A gentle tap may help spread the reagent more evenly.

Reagent Addition

Reagent Removal from Wells

- Place assembled cassette flat on a clean work surface.
- Slightly tilt the cassette while removing the reagent.
- Place the pipette tip on the bottom edge of the well.
- Remove reagents along the side of the well without touching the tissue sections.
- Remove all liquid from the well in each step.

Reagent Removal

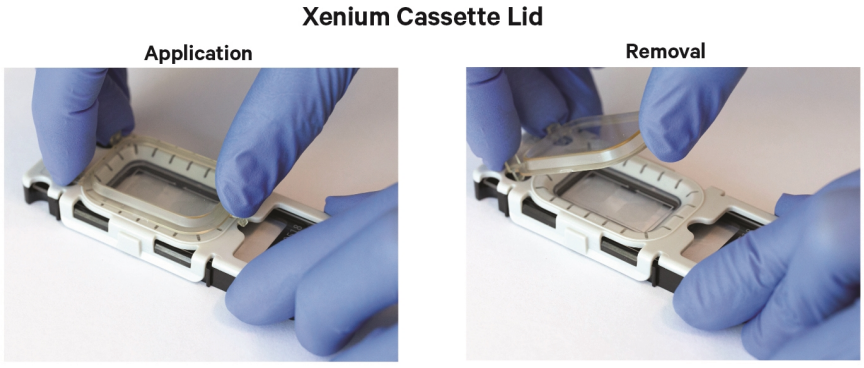
Xenium Cassette Lid Application & Removal

Application

- Place the Xenium Cassette flat on a clean work surface.
- Hold the Xenium Cassette Lid with index and middle finger on two upper tabs and thumb on the lower clip.
- Align the Xenium Cassette Lid with the surface of the Xenium Cassette. Hook the two upper clips into the two holes on the top of the cassette.
- Push the lid down until the lower clip clicks into place.
- Inspect the lid to confirm placement.

Removal

- Place the Xenium Cassette flat on a clean work surface.
- Push on the top of the two upper tabs with index and middle fingers.
- Use thumb to push in on the lower clip.
- While maintaining inward pressure, pull upward with thumb until the lower clip disengages.
- Ensure that no liquid splashes out of the well.

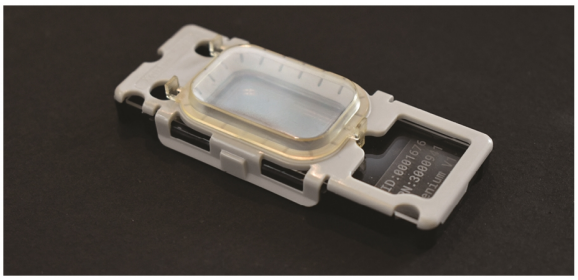


Note that Xenium Cassette Lids are a single use item and should be discarded after each use.

Xenium Cassette Storage

- Store cassettes sealed with a Xenium Cassette Lid at the indicated stopping points listed throughout the protocol and as outlined in the Protocol Steps & Timing on page 14.
- Cassettes should always be stored hydrated with recommended reagent and at 4°C.

Sealed Cassette

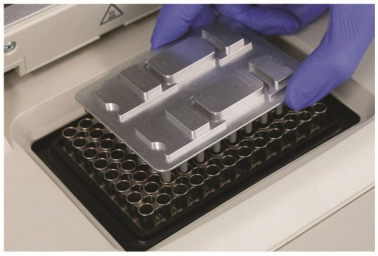


Slide Incubation Guidance

Incubation at a specified temperature

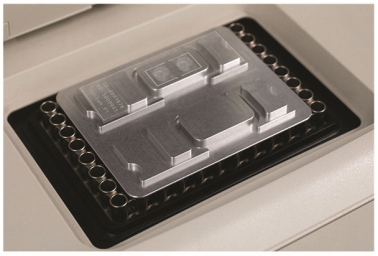
- Position a Xenium Thermocycler Adaptor on a thermal cycler that is set at the incubation temperature. Ensure thermal cycler has reached appropriate temperature prior to starting incubation.

Place Thermocycler Adaptor



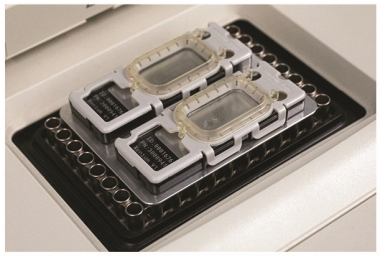
- Ensure that the Thermocycler Adaptor is fully inserted into the thermal cycler and is in contact with the thermal cycler block surface uniformly.
- When incubating a slide, position the slide on the Thermocycler Adaptor with the tissue side facing up.

Incubate Slide



- Ensure the Sample Area is aligned with the corresponding area on the Thermocycler Adaptor. DO NOT close the lid.
- When incubating a slide encased in a cassette, place the assembled unit on the Thermocycler Adaptor with the well facing up. Ensure the cassette is in complete contact with the Thermocycler Adaptor. The cassette should always be sealed with a Xenium Cassette Lid when on the Thermocycler Adaptor unless indicated otherwise.

Incubate Assembled Xenium Cassette



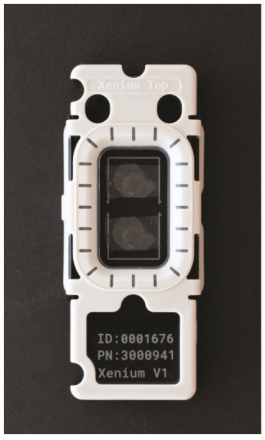
Tightening the thermal cycler lid

- Thermal cycler lid contact with the Xenium Cassette Lid is critical for assay performance.
- Tighten the thermal cycler lid until an audible click is heard.
- Tightening past the click risks breaking the slide.

Incubation at room temperature

- Place the assembled cassette on a flat, clean, non-absorbent work surface.
- Ensure that no absorbent surface is in contact with the reagents on the slide/cassette during incubation.

Cassette Incubation



Tissue Detachment on Xenium Slides

- Monitor section adhesion on Xenium slides throughout the workflow.
- Tissue detachment during the workflow can negatively impact performance. If observed, contact support@10xgenomics.com.
- For more information, refer to Troubleshooting.

Step 1:

Probe Hybridization

1

1.0 Get Started

Each 10x Genomics reagent tube is good for two Xenium slides.

Items			10x PN	Preparation & Handling	Storage
Equilibrate to room temperature					
☐	●	Xenium Probe Dilution Buffer	2000393	Thaw at room temperature. Vortex and centrifuge briefly. Maintain at room temperature after thawing. After use, return to -20°C.	–20°C
☐	●	Xenium Probe Hybridization Buffer	2000390	Thaw at room temperature for 15 min or until completely thawed. Check for precipitate and invert until clear. Maintain at room temperature after thawing. After use, return to -20°C.	–20°C
☐	●	Xenium Human Breast Gene Expression Probes*	2000826	Thaw at room temperature. Preheat by incubating at 95°C for 2 min in a heatblock or waterbath, followed by 1 min on ice. Maintain at room temperature.	–20°C
☐	●	Xenium Mouse Brain Gene Expression Probes*	2000825	Thaw at room temperature. Preheat by incubating at 95°C for 2 min in a heatblock or waterbath, followed by 1 min on ice. Maintain at room temperature.	–20°C
☐	○	Xenium Custom Gene Expression Probes, 50*	3000975	Resuspend custom probes according to the instructions in Probe Hybridization. For freshly resuspended and frozen aliquots, re-equilibrate or thaw at room temperature, respectively. Preheat by incubating at 95°C for 2 min in a heatblock or waterbath, followed by 1 min on ice. Maintain at room temperature.	–20°C
☐	○	Xenium Custom Gene Expression Probes, 100*	3001187	Resuspend custom probes according to the instructions in Probe Hybridization. For freshly resuspended and frozen aliquots, re-equilibrate or thaw at room temperature, respectively. Preheat by incubating at 95°C for 2 min in a heatblock or waterbath, followed by 1 min on ice. Maintain at room temperature.	–20°C
Obtain					
☐		Assembled cassettes	-	Consult Xenium in Situ for FFPE - Deparaffinization & Decrosslinking	-

Items		10x PN	Preparation & Handling	Storage
	containing FFPE or FF tissue samples		(Demonstrated Protocol CG000580) or Xenium in Situ for Fresh Frozen - Fixation & Permeabilization (Demonstrated Protocol CG000581), respectively.	
☐	Nuclease-free Water	-	-	Ambient
☐	10X PBS, pH 7.4	-	-	Ambient
☐	10% Tween-20	-	-	Ambient
☐	Xenium Cassette Lids (16 ct)	3001046	See Tips & Best Practices.	Ambient
☐	Xenium Thermocycler Adaptor	3000954	See Tips & Best Practices.	Ambient

*Thaw appropriate probe panel(s) based on experimental needs. Custom panels are optional.

1.1 Buffer Preparation

Prepare the following buffers fresh before starting the Xenium In Situ Gene Expression workflow. The volumes of each buffer are sufficient for washes in all subsequent steps.

a. Prepare 1X PBS according to the table below before use and maintain at **room temperature.** Add reagents in the order listed. Invert gently to mix.

1X PBS	Stock	Final	1X+10% (µl)	2X+10% (µl)
Nuclease-free Water	-	-	13,500	27,000
10X PBS, pH 7.4	10X	1X	1,500	3,000
Total	**-**	**-**	**15,000**	**30,000**

b. Using 1X PBS from step 1.1a, prepare PBS-Tween Buffer (PBS-T) according to the table below before use and maintain at **room temperature.** Add reagents in the order listed. Invert gently to mix.

PBS-T	Stock	Final	1X+10% (µl)	2X+10% (µl)
1X PBS (prepared at Step 1.1a)	-	-	9,950	19,900
10% Tween-20	10%	0.05%	50	100
Total	**-**	**-**	**10,000**	**20,000**

1.2 Probe Hybridization

Before starting this protocol, ensure that tissue sections have been appropriately deparaffinized and decrosslinked if working with FFPE tissues. Ensure that tissue sections have been appropriately fixed and permeabilized if working with fresh frozen tissues. Consult Xenium in Situ for FFPE - Deparaffinization & Decrosslinking (Demonstrated Protocol CG000580) or Xenium in Situ for Fresh Frozen - Fixation & Permeabilization (Demonstrated Protocol CG000581), respectively, for more information.

TIPS

*During reagent removal steps, ensure that **ALL the liquid is removed** from the wells. See Tips & Best Practices for guidance on Reagent Removal.*

a. Prepare 1X PBS and PBS-T buffers fresh as outlined in step 1.1. The volumes of each buffer are sufficient for washes in all subsequent steps.

b. Prepare a thermal cycler with the following incubation protocol and start the program.

Lid Temperature	Reaction Volume	Run Time
50°C (lid may be turned off if the instrument doesn't enable 50°C)	100 µl	-
Step	**Temperature**	**Time** *hh:mm:ss*
Pre-equilibrate	50°C	Hold
Probe Hybridization	50°C	Overnight (16 - 24 h)
Hold	50°C	Hold

c. Prepare Probe Hybridization Mix according to the tables below. The first table provides preparation instructions for pre-designed probe panels only. The second table provides preparation instructions for custom panels in addition to pre-designed probe panels. Note that custom probes cannot be used in isolation and must be used with pre-designed probe panels.

Probe Hybridization Mix: pre-designed probe panels only

Thaw pre-designed probes at **room temperature**. Remove an aliquot appropriate for the number of desired Xenium slides (see below) and pre-heat/cool according to the Probe Hybridization Get Started table.

Prepare Probe Hybridation Mix shortly before use and maintain at **room temperature**. Add reagents in the order listed. Pipette mix and centrifuge briefly.

	Probe Hybridization Mix *(pre-designed probe panels only)*	**10x PN**	**1X+10% (µl)**	**2X+10% (µl)**
●	Xenium Probe Hybridization Buffer	2000390	330.0	660.0
●	Xenium Probe Dilution Buffer	2000393	185.6	371.2
●	Xenium Human Breast Gene Expression Probes or Xenium Mouse Brain Gene Expression Probes	2000826 2000825	34.4	68.8
	Total	**-**	**550.0**	**1,100.0**

Probe Hybridization Mix: pre-designed probe panels with custom probe panels

Custom probes are delivered lyophilized and must be resuspended before use according to the following instructions:

- Centrifuge custom probe panel tube briefly
- Resuspend in **625 µl** of **room temperature** Xenium Probe Dilution Buffer
- Replace the cap firmly and agitate for **5 min**
- Centrifuge briefly and maintain at **room temperature**

If custom probes are already resuspended, thaw at **room temperature**. For both freshly resuspended and thawed custom probe panels, remove an aliquot appropriate for the number of desired Xenium slides (see below) and pre-heat/cool according to the Probe Hybridization Get Started table.

Prepare Probe Hybridation Mix shortly before use and maintain at **room temperature**. Add reagents in the order listed. Pipette mix and centrifuge briefly.

	Probe Hybridization Mix *(pre-designed probe panels with custom probe panels)*	**10x PN**	**1X+10% (µl)**	**2X+10% (µl)**
●	Xenium Probe Hybridization Buffer	2000390	330.0	660.0
●	Xenium Probe Dilution Buffer	2000393	151.2	302.4
●	Xenium Human Breast Gene Expression Probes or Xenium Mouse Brain Gene Expression Probes	2000826 2000825	34.4	68.8
○	Xenium Custom Gene Expression Probes, 50 or Xenium Custom Gene Expression Probes, 100	3000975 3001187	34.4	68.8
	Total	**-**	**550.0**	**1,100.0**

TIPS

Record the Custom Panel Design ID and Slide Number before starting workflow. This information is critical for identifying the correct electronic decode file when setting up the Xenium Analyzer in downstream steps.

d. Retrieve the assembled Xenium Cassette containing FFPE or fresh frozen tissue sections.

e. Remove all PBS-T from FFPE or fresh frozen tissues as prepared according to Xenium in Situ for FFPE - Deparaffinization & Decrosslinking (CG000580) or Xenium in Situ for Fresh Frozen - Fixation & Permeabilization (CG000581) Demonstrated Protocols, respectively.

f. Add **500 µl** room-temperature Probe Hybridization Mix along the side of the well to uniformly cover the tissue section(s), without introducing bubbles.

g. Apply a new Xenium Cassette Lid on the Xenium Cassette and place on the Xenium Thermocycler Adaptor on the pre-heated thermal cycler. Tightly close the thermal cycler lid until an audible click is heard.

h. Skip Pre-equilibrate step to initiate Probe Hybridization.

i. After Probe Hybridization is complete, **immediately** proceed to next step.

Step 2:

Post Hybridization Wash

2

2.0 Get Started

Each 10x Genomics reagent tube is good for two Xenium slides.

Items			10x PN	Preparation & Handling	Storage
Equilibrate to room temperature					
☐	◯	Xenium Post Hybridization Wash Buffer	2000395	Thaw at room temperature for 30 min or until thawed completely. Vortex and centrifuge briefly. Keep the buffer at room temperature after thawing. After use, return to -20°C.	–20°C
Obtain					
☐		PBS-T	-	Prepared at Step 1.1.	Ambient

2.1 Post Hybridization Wash

a. Remove the Xenium Cassette from the Thermocycler Adaptor and place on a flat, clean work surface. DO NOT let the cassette cool down before proceeding to PBS-T washes.

Fluid on the Thermocycler Adaptor may indicate a reagent leak from the cassette. See Troubleshooting for more details.

b. Remove the Xenium Cassette Lid and using a pipette, remove all Probe Hybridization Mix from well corners. Discard old Cassette Lids.

If processing two slides at a time, remove and add reagent from first slide before proceeding to second slide. Ensure tissue sections are always covered with reagent in between removal and addition steps to avoid drying out of tissue samples.

c. **Immediately** add **500 µl** PBS-T prepared at step 1.1 along the side of the well to uniformly cover the tissue sections, without introducing bubbles. Removal and addition of buffers should be done quickly to prevent drying of tissue sections.

Small bubbles on the surface of the slide are normal and unlikely to compromise assay performance. DO NOT aspirate or pop bubbles, as this can lead to detachment or scratching of the tissue.

d. Incubate for **1 min** at **room temperature**.

e. Prepare a thermal cycler with the following incubation protocol and start the protocol.

Lid Temperature	Reaction Volume	Run Time
37°C (lid may be turned off if the instrument doesn't enable 37°C)	100 µl	-
Step	**Temperature**	**Time** *hh:mm:ss*
Pre-equilibrate	37°C	Hold
Post Hybridization Wash	37°C	00:30:00
Hold	37°C	Hold

f. Using a pipette, remove all PBS-T from well corners.

g. Add **500 µl** PBS-T.

h. Incubate for **1 min** at **room temperature**.

i. Remove all PBS-T.

j. Add **500 µl** Xenium Post Hybridization Wash Buffer to the well.

k. Apply a new Xenium Cassette Lid on the Xenium Cassette and place on the Thermocycler Adaptor on the pre-heated thermal cycler. Close the thermal cycler lid.

l. Skip Pre-equilibrate step to initiate Post Hybridization Wash.

Start thawing Ligation reagents during Post Hybridization Wash incubation as outlined in the Get Started table in step 3.0.

m. After the Post Hybridization Wash is complete, **immediately** proceed to the next step.

Step 3:

Ligation

3.0 Get Started

Each 10x Genomics reagent tube is good for two Xenium slides.

Items		10x PN	Preparation & Handling	Storage
Equilibrate to room temperature				
☐	Xenium Ligation Buffer	2000391	Thaw at room temperature for 15 min or until completely thawed. Vortex and centrifuge briefly. Maintain at room temperature after thawing.	–20°C
Place on ice				
☐	Xenium Ligation Enzyme A	2000397	Pipette mix and centrifuge briefly. Maintain on ice until ready to use.	–20°C
☐	Xenium Ligation Enzyme B	2000398	Pipette mix and centrifuge briefly. Maintain on ice until ready to use.	–20°C
Obtain				
☐	PBS-T	-	Prepared at Step 1.1.	Ambient

3.1 Ligation

a. Prepare Ligation Mix shortly before using. Add reagents in the order listed. Pipette mix 10X and centrifuge briefly. Maintain on ice.

	Ligation Mix	10x PN	1X+10% (µl)	2X+10% (µl)
●	Xenium Ligation Buffer	2000391	481.2	962.5
●	Xenium Ligation Enzyme A	2000397	13.8	27.5
●	Xenium Ligation Enzyme B	2000398	55.0	110.0
	Total	**-**	**550.0**	**1,100.0**

b. Remove the Xenium Cassette from the Thermocycler Adaptor and place on a flat, clean work surface.

c. Remove the Xenium Cassette Lid and using a pipette, remove all Xenium Post Hybridization Wash Buffer from the well. Discard old Cassette Lids.

If processing two slides at a time, remove and add reagent from first slide before proceeding to second slide. Ensure tissue sections are always covered with reagent in between removal and addition steps to avoid drying out of tissue samples.

d. **Immediately** add **500 µl** PBS-T prepared at step 1.1 to the well. Removal and addition of buffers should be done quickly.

e. Incubate at **room temperature** for **1 min**.

f. Prepare a thermal cycler with the following incubation protocol. Place a Thermocycler Adaptor on the thermal cycler and start the program.

Lid Temperature	Reaction Volume	Run Time
37°C (lid may be turned off if the instrument doesn't enable 37°C)	100 µl	-
Step	**Temperature**	**Time** *hh:mm:ss*
Pre-equilibrate	37°C	Hold
Ligation	37°C	02:00:00

g. Using a pipette, remove all PBS-T from well corners.

h. Add **500 µl** PBS-T.

i. Incubate at **room temperature** for **1 min**.

j. Remove all PBS-T.

k. **Repeat** steps h-j one more time.

l. Add **500 µl** Ligation Mix to the well.

m. Apply a new Xenium Cassette Lid on the Xenium Cassette and place on the Thermocycler Adaptor on the pre-heated thermal cycler. Close the thermal cycler lid.

n. Skip Pre-equilibrate step to initiate Ligation.

TIPS

Start thawing Amplification reagents during Ligation incubation as outlined in the Get Started table in step 4.0.

o. After Ligation is complete, **immediately** proceed to next step.

Step 4:

Amplification

4

4.0 Get Started

Each 10x Genomics reagent tube is good for two Xenium slides.

Item		10x PN	Preparation & Handling	Storage
Place on ice				
☐ ●	Xenium Amplification Mix	2000392	Thaw on ice. Vortex and centrifuge briefly.	–20°C
☐ ●	Xenium Amplification Enyzme	2000399	Pipette mix and centrifuge briefly. Maintain on ice until ready to use.	–20°C
Obtain				
☐	PBS-T	-	Prepared at Step 1.1.	Ambient
☐	TE Buffer, TRIS-EDTA, 1X Solution, pH 8.0	-	-	Ambient

4.1 Amplification

a. Prepare Amplification Master Mix shortly before use. Add reagents in the order listed. Pipette mix 10X and centrifuge briefly. Maintain on ice.

	Amplification Master Mix	10x PN	1X +10% (µl)	2X +10% (µl)
●	Xenium Amplification Mix	2000392	495.0	990.0
●	Xenium Amplification Enzyme	2000399	55.0	110.0
	Total	**-**	**550.0**	**1,100.0**

b. Remove the Xenium Cassette from the Thermocycler Adaptor and place on a flat, clean work surface.

c. Remove the Xenium Cassette Lid and using a pipette, remove all Ligation Mix from the well. Discard old Cassette Lids.

If processing two slides at a time, remove and add reagent from first slide before proceeding to second slide. Ensure tissue sections are always covered with reagent in between removal and addition steps to avoid drying out of tissue samples.

d. Add **500 µl** PBS-T prepared at step 1.1 to the well.

e. Incubate for **1 min** at **room temperature**.

f. Prepare a thermal cycler with the following incubation protocol. Place a Thermocycler Adaptor on the thermal cycler and start the program.

Lid Temperature	Reaction Volume	Run Time
30°C (lid may be turned off if the instrument doesn't enable 30°C)	100 µl	-
Step	**Temperature**	**Time** *hh:mm:ss*
Pre-equilibrate	30°C	Hold
Amplification	30°C	02:00:00

g. Using a pipette, remove all PBS-T from well corners.

h. Add **500 µl** PBS-T.

i. Incubate for **1 min** at **room temperature**.

j. Remove all PBS-T.

k. **Repeat** steps h-j one more time.

l. **Immediately** add **500 µl** Amplification Master Mix to the well.

m. Apply a new Xenium Cassette Lid on the Xenium Cassette and place on the Thermocycler Adaptor on the thermal cycler. Close the thermal cycler lid.

n. Skip pre-equilibrate step to initiate Amplification.

TIPS

Start thawing Autofluorescence Quenching reagents during Amplification incubation as outlined in the Get Started table in step 5.0.

o. After Amplification is complete, **immediately** proceed to next step.

4.2 Post Amplification Wash

a. Remove the Xenium Cassette from the Thermocycler Adaptor and place on a flat, clean work surface.

b. Remove the Xenium Cassette Lid and using a pipette, remove all Amplification Mix from the well. Discard old Cassette Lids.

c. Add **500 µl** TE Buffer to the well.

d. Incubate **1 min** at **room temperature**.

e. Remove all TE buffer.

f. **Repeat** steps c-e one more time.

g. Add **500 µl** TE Buffer to the well.

h. Proceed to next step or apply Xenium Cassette Lid on the Xenium Cassette and store the slides in TE Buffer at **4°C overnight or ≤4 days**. DO NOT remove the Xenium Cassette Lid during storage.

Step 5:

Autofluorescence Quenching

5

5.0 Get Started

Each 10x Genomics reagent tube is good for two Xenium slides.

Items			10x PN	Preparation & Handling	Storage
Equilibrate to room temperature					
☐	●	Xenium Autofluorescence Mix	2000753	Thaw in a thermomixer (with 2.0-ml thermoblock) for 15 min at 37°C, 300 rpm with shaking. Cool to room temperature for 5 min. Vortex for 30 sec and centrifuge briefly. Alternatively, thaw in a waterbath for 15 min at 37°C. Cool to room temperature for 5 min. Vortex for 30 sec and centrifuge briefly.*	–20°C
☐	○	Reducing Agent B	2000087	Thaw at room temperature. Vortex and centrifuge briefly.	–20°C
☐	●	Xenium Nuclei Staining Buffer	2000762	Thaw at room temperature. Vortex and centrifuge briefly. Keep in the dark until ready to use.	–20°C
Obtain					
☐		Nuclease-free Water	-	-	Ambient
☐		1X PBS	-	Prepared at Step 1.1.	Ambient
☐		PBS-T	-	Prepared at Step 1.1.	Ambient
☐		100% Ethanol	-	-	Ambient

TIPS

*Pre-heat thermomixer or waterbath to 37°C in advance of intended use.

5.1 Autofluorescence Quenching

a. Prepare the following for Autofluorescence Quenching:

i. Prepare diluted Reducing Agent B. Add reagents in the order listed. Maintain at room temperature.

	Diluted Reducing Agent B	10x PN	Stock	Final	1X+10% (µl)	2X+10% (µl)
	1X PBS (prepared at Step 1.1)	-	-	-	544.5	1,089.0
○	Reducing Agent B	2000087	-	-	5.5	11.0
	Total	**-**	-	-	**550.0**	**1,100.0**

ii. Prepare 70% Ethanol. Add reagents in the order listed. Maintain at room temperature.

70% Ethanol	10x PN	Stock	Final	1X+10% (µl)	2X+10% (µl)
Nuclease-free Water	-	-	-	330.0	660.0
100% Ethanol	-	100%	70%	770.0	1,540.0
Total	**-**	-	-	**1,100.0**	**2,200.0**

iii. Prepare Autofluorescence Solution using thawed Xenium Autofluorescence Mix prepared according to step 5.0. Add reagents in the order listed and vortex to mix. Maintain at room temperature in the dark until ready to use.

	Autofluorescence Solution	10x PN	Stock	Final	1X+10% (µl)	2X+10% (µl)
	100% Ethanol	-	100%	-	544.5	1,089.0
●	Xenium Autofluorescence Mix	2000753	-	-	5.5	11.0
	Total	**-**	-	-	**550.0**	**1,100.0**

b. Retrieve the Xenium Cassette from step 4.2h and place on a flat, clean work surface.

c. Remove the Xenium Cassette Lid and using a pipette, remove all TE Buffer from the well. Discard old Cassette Lids.

If processing two slides at a time, remove and add reagent from first slide before proceeding to second slide. Ensure tissue sections are always covered with reagent in between removal and addition steps to avoid drying out of tissue samples.

d. Add **1,000 µl** 1X PBS prepared at step 1.1 to the well and incubate for **1 min** at **room temperature**.

e. Remove all 1X PBS.

f. **Repeat** steps d-e two more times.

g. Add **500 µl** Diluted Reducing Agent B prepared at step 5.1ai to the well.

h. Apply a new Xenium Cassette Lid on the Xenium Cassette, and incubate for **10 min** at **room temperature**.

i. Remove the Xenium Cassette Lid and using a pipette, remove all Diluted Reducing Agent B from the well. Discard old Cassette Lids.

j. Add **1,000 µl** 70% Ethanol prepared at step 5.1aii. Wait **1 min**.

k. Remove all 70% Ethanol.

l. Add **1,000 µl** 100% Ethanol. Wait **1 min**.

m. Remove all 100% Ethanol.

n. **Repeat** steps l-m for a total of two washes.

o. **Immediately** add **500 µl** Autofluorescence Solution prepared at step 5.1aiii. Pipette mix thoroughly before dispensing onto sample to prevent settling of reagent.

p. Apply a new Xenium Cassette Lid on the Xenium Cassette, and incubate for **10 min** at **room temperature in the dark**.

q. Prepare a thermal cycler with the following incubation protocol. Place a Thermocycler Adaptor on the thermal cycler and start the program.

Lid Temperature	Reaction Volume	Run Time
37°C (lid may be turned off if the instrument doesn't enable 37°C)	100 µl	-
Step	**Temperature**	**Time** *hh:mm:ss*
Pre-equilibrate	37°C	Hold
Drying	37°C	00:05:00

r. Remove the Xenium Cassette Lid and using a pipette, remove all Autofluorescence Solution. Discard old Cassette Lids.

s. Add **1,000 µl** 100% Ethanol. Wait **2 min**.

t. Remove all 100% Ethanol.

u. **Repeat** steps s-t two more times.

v. Place Xenium Cassette **without lid** on the Thermocycler Adaptor on the thermal cycler to dry. DO NOT close the thermal cycler lid.

w. Skip pre-equilibrate step to initiate Drying.

x. **Immediately** remove the Xenium Cassette from the Thermocycler Adaptor and place on a flat, clean work surface.

y. Add **1,000 µl** 1X PBS prepared at step 1.1 to rehydrate the tissue and incubate for **1 min** at **room temperature in the dark**.

z. Remove all 1X PBS.

aa. Add **1,000 µl** PBS-T and incubate for **2 min** at **room temperature in the dark**.

Optional: photograph the slide against a white background. This image can be used for comparison purposes to identify tissue detachment downstream in the workflow. See Troubleshooting for more details.

ab. Proceed to next step (recommended), or for long-term storage (≤4 days), apply Xenium Cassette Lid on the Xenium Cassette and store slides at **4°C in the dark**. DO NOT remove the Xenium Cassette Lid during storage.

5.2 Nuclei Staining

a. Retrieve thawed Xenium Nuclei Staining Buffer prepared according to the Get Started table in step 5.0.

b. Remove all PBS-T.

c. Add **500 µl** Xenium Nuclei Staining Buffer and incubate **1 min** at **room temperature in the dark**.

d. Remove all Nuclei Staining Buffer.

e. Add **1,000 µl** PBS-T prepared at step 1.1 to the well.

f. Incubate for **1 min** at **room temperature in the dark**.

g. Remove all PBS-T.

h. **Repeat** steps e-g two more times.

i. Add **1,000 µl** PBS-T.

j. Apply a Xenium Cassette Lid on the Xenium Cassette. Proceed to the Xenium Analyzer User Guide with Readiness Test (CG000584), or for long-term storage (≤4 days), store slides at **4°C in the dark**. DO NOT remove the Xenium Cassette Lid during storage.

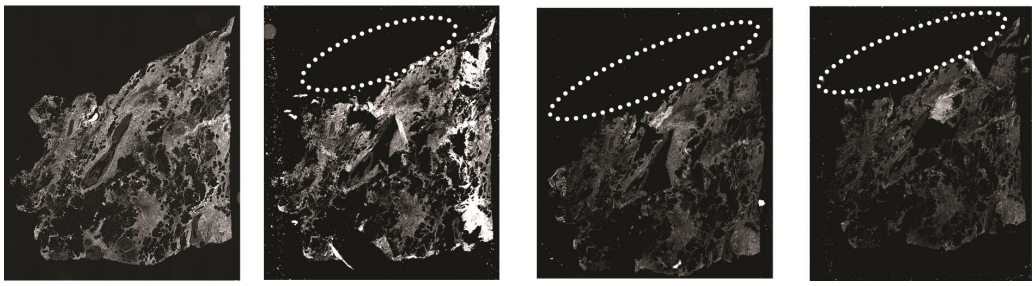
Troubleshooting

Tissue Detachment and Folding

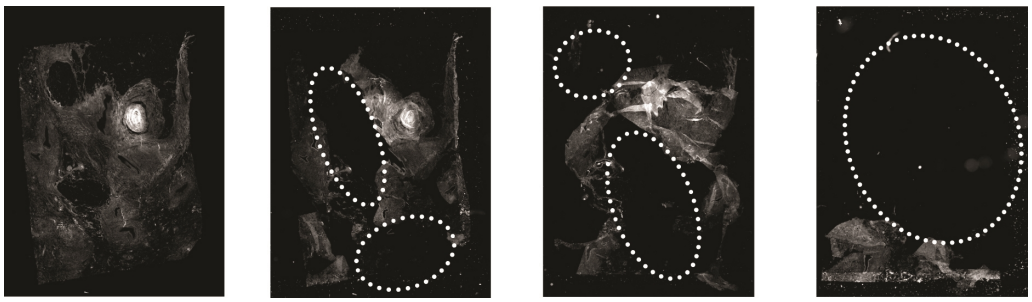
Tissue detachment may result in a lack of decodable data in the region where detachment occurred. If the tissue has folded on itself, this may also cause elevated signal in the overlapping areas. Inspect images carefully to identify these areas. If tissue detachment is observed during this workflow, contact support@10xgenomics.com

Tissue Detachment in Human Breast as viewed on Xenium Analyzer Overview Scan

No Detachment	10-25% Detachment		
	Section 1	Section 2	Section 3



No Detachment	70-95% Detachment		
	Section 1	Section 2	Section 3



Percentages represent tissue detachment/"area that cannot be analyzed" at the end of the Xenium Analyzer workflow. White circles indicate areas of tissue detachment.

Tissue Detachment on Xenium Slides

Tissue sections may detach from Xenium slides during on-slide workflows. Tissue adhesion to the slide is impacted both by tissue processing and tissue quality.

Careful tissue preparation is critical for adhesion in on-slide workflows. Consult Xenium In Situ for Fresh Frozen - Tissue Preparation Guide (Document CG000578) and Xenium In Situ for FFPE - Tissue Preparation Guide (Document CG000579) for tissue QC and sectioning best practices. Below are some additional best practices for minimizing detachment during on-slide workflows:

- Do not pipette directly onto the tissue.
- Gently add and remove reagents from the well. Forceful addition or removal of reagents can agitate tissue and lead to detachment.
- Avoid touching in and around the Sample Area of the Xenium slide.
- Work quickly and carefully during reagent addition and removal.

In addition to following best practices, it is possible to monitor section adhesion on Xenium slides throughout the workflow. Taking a photograph of the slide at the beginning of the on-slide workflow and comparing with post-assay workflow images can help identify whether tissue shape has changed significantly, an indication of detachment. Steps when slide photos can be taken are noted in the protocol. These QC images can be compared with the DAPI overview scan as part of the Web Summary file to see whether tissue morphology has changed in the workflow.

If tissue detachment occurs, send pictures to support@10xgenomics.com for further assistance.

Bubbles during Workflow

Bubbles may occur throughout the Xenium In Situ Gene Expression workflow, including after Probe Hybridization and Ligation, and during PBS-T washes. Bubbles floating on the surface of the slide are unlikely to compromise assay performance. However, bubbles that are in contact with the tissue during a Xenium Analyzer run may result in a lack of decodable data in the tissue area where the bubbles occurred.

Avoid generating bubbles during reagent dispensing by pipetting slowly and avoiding expelling air from the pipette tip. Gently tap or rock the cassette after reagent dispension and inspect the cassette carefully to ensure liquid is fully covering the tissue. DO NOT aspirate or pop the bubbles as this could lead to tissue detachment or scratching of the tissue. Ensure there are no bubbles on the assembled cassette before loading it into the Xenium Analyzer.

Number of Washes

Post Hybridization and post-Ligation washes are critical for assay performance. Failure to perform the correct number of washes can reduce the fraction of usable decodable data. A similar effect is observed when washing for less than the recommended time, or when reagent is carried over during the washes. Remove all liquid from the well when washing, and refer to User Guide for correct number of washes and incubation times.

Samples Dry Out

Drying of tissue samples may lead to decreased decoding efficiency and unusable data. Work quickly and ensure reagents are dispensed evenly across tissues during incubation and wash steps throughout the workflow to prevent drying out of tissues. If processing two slides at a time, remove and add reagent from first slide before proceeding to second slide. Ensure tissue sections are always covered with reagent in between removal and addition steps. Note that there are no safe stopping points except for those described in the protocol and outlined specifically in the Protocol Steps & Timing.

Cassette Assembly Failure

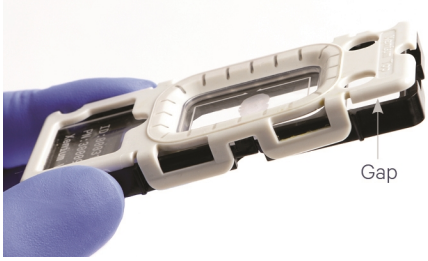
Incorrect assembly of the Xenium Cassette with a Xenium slide can negatively impact assay performance. Always dry the front and the back of the slide completely using a lint-free laboratory wipe while avoiding touching or damaging of the tissue sections. Inspect the slide carefully to ensure it is seated fully within the cassette before assembly.

Example scenarios that may indicate incorrect Xenium Cassette assembly are described below:

- If a gap appears between the two halves of the cassette after assembly.
- If the cassette does not click shut or appears domed after assembly.
- If the Xenium Thermocycler Adaptor is wet following removal from the thermal cycler, indicating reagent leakage from the cassette.

If the cassette is incorrectly assembled, disassemble and reassemble the cassette as instructed in the following pages.

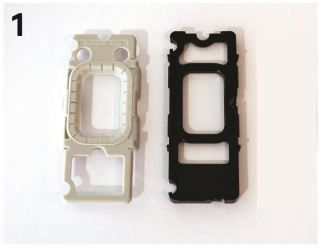
Incorrect cassette assembly as indicated by a gap between the two halves of the cassette

Correct cassette assembly	Incorrect cassette assembly
	

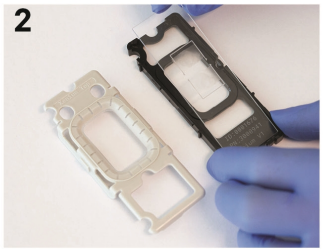
Xenium Cassette Assembly

Exercise caution when handling slide edges to prevent injury.

Place top and bottom halves of cassette on bench

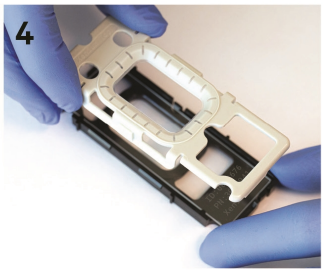


Place Xenium slide with tissue side facing upwards into bottom half of cassette; ensure label is toward bottom of cassette

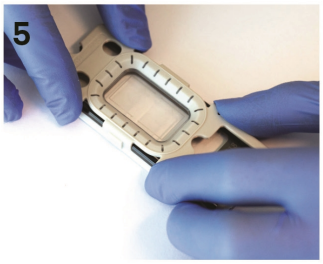


Press slide down into grooves of the bottom half of the cassette until it sits firmly in place

Secure clips of top half with tabs of bottom half (on both sides)



Apply even pressure on top of cassette until all clips click shut. Verify that clips are completely secured over tabs

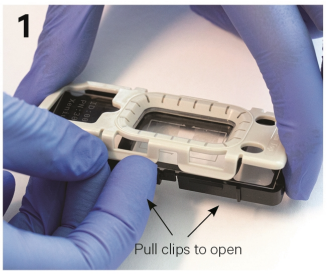


Slides in images are representative.

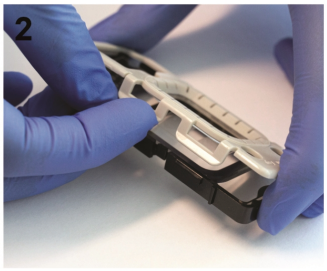
TIPS Once cassette is assembled, DO NOT remove slide until after Xenium Analyzer imaging and decoding for optional H&E staining step.

Xenium Cassette Removal

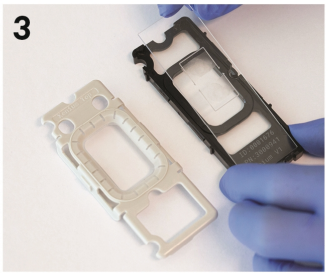
Pull inner clips from inner tabs to detach top and bottom halves of cassette



Open cassette by continuing to lift inner clips upward



Hold slide by the label and lift slide out from bottom half



Slides in images are representative.

Xenium Cassette Lid Cleaning

Xenium Cassette Lids are a single use item and are discarded following reagent incubations as indicated throughout the User Guide. Cassette lids that are accidentally dropped may be reused after thorough cleaning. The following cleaning procedure should be performed in between every step of the workflow where the lid is used (following overnight probe hybridization, ligation, etc.). Note that PBS-T washes DO NOT require sealing of the cassette.

Cleaning Procedure:

- Rinse the lid under running MilliQ water
- Spray with 70% isopropanol
- Rinse under running MilliQ water
- Spray with 70% isopropanol a second time
- Rinse under running MilliQ water
- Air dry

Incorrect Autofluorescence Quenching

Variation in stain color is normal and tissue-type dependent in tissue sections correctly stained with Autofluorescence Solution. Incorrect staining scenarios are listed below:

- Uneven staining with Autofluorescence Solution may be visible as a non-uniform stain across a tissue section.
- Overquenching can cause tissue to overheat on the Xenium Analyzer, and data generated in the overheated spots may be compromised or missing.
- If no Autofluorescence Quenching is performed, the Xenium In Situ Gene Expression workflow will need to be repeated.

Ensure Autofluorescence Solution is well mixed and dispensed uniformly across the tissue sections to avoid uneven staining. Cassette should be sealed properly and firmly during incubation to prevent reagent evaporation.

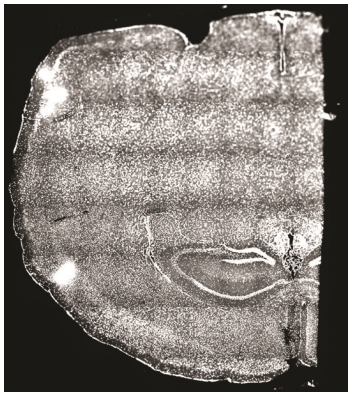
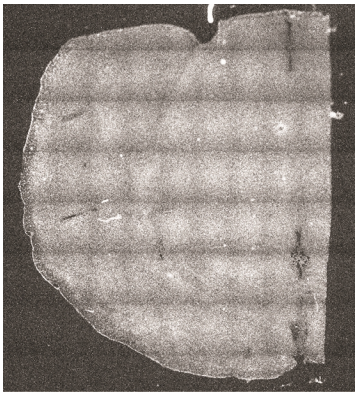
Incorrect or insufficient Autofluorescence Quenching may cause variation in staining of tissues *(Mouse Brain pictured below)*

Unstained	Normally Stained	Overstained	Understained
			

Incorrect Nuclei Staining

Incorrect staining of nuclei may lead to poor image quality and an inability to easily identify tissue or regions of interest when selecting areas to image during a Xenium Analyzer overview scan. Follow the Nuclei Staining protocol as instructed using the Xenium Nuclei Staining Buffer provided in the Xenium Slides & Sample Prep Reagents Kit - (2 slides, 2 rxns), PN-1000460. Confirm Xenium Nuclei Staining Buffer is well mixed and applied uniformly across tissue sections. All incubations with Xenium Nuclei Staining Buffer should be performed in the dark. If an alternate staining protocol or buffer is used, lower quality images may be obtained.

Incorrect or insufficient nuclei staining may impact image quality and region of interest selection *(as viewed on a Xenium Analyzer overview scan)*

Correct Nuclei Staining Protocol	No Nuclei Staining Performed
	

Appendix

Probe Panel Selection

Ensure that a compatible gene panel has been selected prior to executing the Xenium In Situ Gene Expression workflow. 10x Genomics provides the option of using pre-designed gene panels. Additionally, pre-designed panels may be customized by adding genes of interest.

Pre-designed Gene Panels

a. Xenium Human Breast Gene Expression, 2 rxns, PN-1000462 (248 genes)

b. Xenium Mouse Brain Gene Expression, 2 rxns, PN-1000463 (280 genes)

Custom Gene Panels

Contact 10x Genomics via email at customerservice@10xgenomics.com for information about designing custom gene panels that are compatible with pre-designed panels.

If utilizing a custom panel, the Design ID on the label of the tube containing the custom panel should match with the first portion of the custom gene panel electronic file name.

Sample Shipping

Processed Xenium slides may be shipped following the Xenium In Situ Gene Expression workflow. After Nuclei Staining, remove all PBS-T from last step, dissassemble the Xenium Cassette, and place slides in a mailer filled to capacity with PBS-T. Ship the slide mailer containing processed Xenium slides in a container with ice packs. Place no more than two slides per mailer. Note that assay performance may be compromised post-shipping and handling.